Plasma Antiprotease Status in Different Respiratory Disorders

R Rai, M Phadke

Citation

R Rai, M Phadke. *Plasma Antiprotease Status in Different Respiratory Disorders*. The Internet Journal of Pulmonary Medicine. 2006 Volume 7 Number 1.

Abstract

In the present study we estimated plasma concentration and activity of antiproteases in control group and in the study groups of patients with chronic obstructive pulmonary disease (COPD), emphysema, bronchiectasis and bronchial asthma. \mathbb{I}_1 -proteinase inhibitor (\mathbb{I}_1 -PI) concentration was significantly increased in all study groups as compared to control group. In patients with emphysema, bronchiectasis, and bronchial asthma. \mathbb{I}_2 -macroglobulin (\mathbb{I}_2 -M) concentration was significantly increased as compared to control group. But in patients with COPD, \mathbb{I}_2 -M concentration was comparable to that of control group. Antitryptic and antielastase activity in condition of COPD was found to increase significantly. But in the other three study groups antitryptic activity was found to decrease. Antielastase activity was not found to vary significantly in bronchiectasis, and was found to decrease significantly in emphysema and bronchial asthma. Significant correlation between antiproteases concentration, antiproteases activity and pulmonary function tests was found only in some study groups.

INTRODUCTION

The lung encounters most of the toxins, particles and infectious agents in our environment before any other organ does₁. Cigarette smoking, excessive inhalation of polluted air, and respiratory infections result in lung irritation and the migration of phagocytic cells to these areas of stress. A major component released from human neutrophil granules during the process of phagocytosis is a proteinase referred to as human neutrophil elastase. This enzyme has been shown to catalyze the degradation of all of the major connective tissue components - including elastin, collagen, and proteoglycan and it is believed to be primarily responsible for the destruction of lung alveoli associated with the development of pulmonary emphysema₂. Normally, the lung is adequately protected against proteases by antiproteases, proteins that rapidly bind to proteases, thereby irreversibly inhibiting their proteolytic activity, \mathbb{I}_1 -proteinase inhibitor (\mathbb{I}_1 -PI), secretory leukoprotease inhibitor (SLPI), and I₂macroglobulin (1,-M) are antiproteases central to the pulmonary antiproteolytic defenses₃.

 \mathbb{I}_1 -PI is an important plasma proteinase inhibitor (52 kDa) and is synthesized by hepatocytes and macrophages₄. \mathbb{I}_1 -PI acts against trypsin, chymotrypsin, plasmin and possibly thrombin, but the inhibition of greatest clinical significance is against neutrophil elastase and collagenase. \mathbb{I}_2 -M is a large glycoprotein (720 kDa) of plasma and is synthesized by a variety of cell types, including monocytes, hepatocytes and astrocytes₄. \mathbb{I}_2 -M inactivates endopeptidases from all four classes (seryl, cysteinyl, aspartyl, metallo). \mathbb{I}_2 -M is an effective inhibitor of human neutrophil elastase , however it is less efficient than \mathbb{I}_1 -PI₅.

Proteases-antiproteases balance is essential for the normal lung function. Various investigators have studied the protease-antiprotease balance in the development of different respiratory conditions like COPD, emphysema, bronchiectasis and bronchial asthma_{6,7,8}. COPD is characterized by airflow limitation that is not fully reversible. The airflow limitation is usually both progressive and associated with an abnormal inflammatory response of the lungs to noxious particles or gases₉. Emphysema is a condition of the lung characterized by abnormal permanent enlargement of the air spaces distal to the terminal bronchiole, accompanied by destruction of their walls₁₀. Bronchiectasis is the chronic abnormal dilatation and distortion of bronchi caused by destruction of the elastic and muscular component of the bronchial wall₁₀. Asthma is an inflammatory disease of the airways in which the mucous membrane and layers of the bronchi become thickened and the mucous glands enlarge, reducing airflow in the lower respiratory tract₁₁.

Pulmonary function tests are widely used in the evaluation and management of patients with known or suspected disorders of respiration. The volumes of air moving in and out of the lungs and remaining in them are of great importance. Forced vital capacity (FVC) is the maximum volume of air which can be breathed out as forcefully and rapidly as possible following a maximum inspiration. Clinical significance of FVC is to distinguish between restrictive and obstructive lung disorders. FEV₁ is the volume of air expired in 1st second of exhalation₁₂.

The aim of the present work was to study and estimate plasma concentration and activity of antiproteases in control group and in the study groups of patients with COPD, emphysema, bronchiectasis and bronchial asthma, and to find out correlation between these estimations and pulmonary functions of patients, if any.

MATERIALS AND METHODS

All chemicals were of the highest purity available and used as received. I₁-PI concentrations and I₂-M concentrations were estimated by using commercial kits and were purchased from Spinreact, Spain. Plasma total protein was determined by using commercial kit and was purchased from Quali Test, Rashmi Diagnostic Pvt. Ltd. Porcine pancreatic elastase (EC 3.4.21.36) and N-Succinyl-Ala-Ala-Ala pnitroanilide were purchased from Sigma Chemical Co., St. Louis, MO. Bovine trypsin was purchased from, S.D. finechem Pvt. Ltd. Boisar. I-N-benzoyl-DL-arginine-pnitroanilide (BAPNA) was purchased from Fluka. Sodium chloride (NaCl), Tris (hydroxymethyl) Aminomethane (tris buffer) were purchased from Qualigens Fine Chemicals Mumbai. All the other chemicals were of analytical reagent grade.

The patients included in the study were from respiratory medicine OPD of Lokmanya Tilak Municipal Medical College and General Hospital Sion, Mumbai.

The study group included 111 patients. Further patients were subgrouped depending upon the diagnosis as : COPD = 24, Emphysema = 20, Bronchiectasis = 32 and Bronchial asthma = 35. The control group included 50 healthy, non-smokers who had no history of lung disease and had normal pulmonary function tests.

Morning venous blood samples were collected for the study of various parameters and taken in EDTA containers. The plasma collected by centrifugation for 10 min. was either tested immediately or frozen and stored at -25° C.

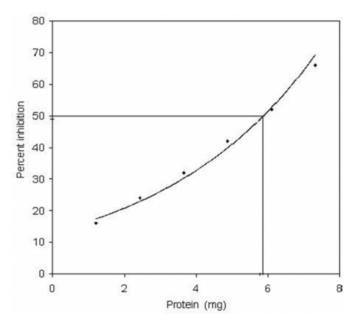
The various parameters estimated were, Plasma \mathbb{I}_1 -PI concentration, Plasma \mathbb{I}_2 -M concentration, Antitryptic activity and Antielastase activity.

The determination of \mathbb{I}_1 -PI concentration and \mathbb{I}_2 -M concentration was based on the reaction between \mathbb{I}_1 -PI and \mathbb{I}_2 -M as antigen and the specific antiserum as antibody. This reaction forms an insoluble complex producing a turbidity which is measured spectrophotometrically at 340 nm. The results were evaluated using a reference curve prepared with the aid of calibrator dilutions.

Antitryptic activity was determined by incubating trypsin and its substrate BAPNA with different volumes of plasma and without plasma and estimating the mg of plasma protein required to inhibit the enzyme to the extent of 50% at optimum pH and optimum temperature condition. The antitryptic inhibitors of plasma inhibit the hydrolysis of \mathbb{I} -Nbenzoyl-DL-arginine-p-nitroanilide (BAPNA) by trypsin in tris buffer, pH 8.0. The reaction is stopped by adding acetic acid (30% v/v) and the absorbance of p-nitroaniline, liberated after hydrolysis is read at 410 nm. The percent inhibition of trypsin was calculated by comparing with positive control (without plasma) which is taken as 100% activity. The mg of protein required for 50% inhibition was found from the plot of the protein concentration (mg) against percent inhibition₁₂₂₁₃ (Figure 1).

Figure 1

Figure 1 : Antitryptic activity of plasma.

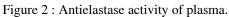


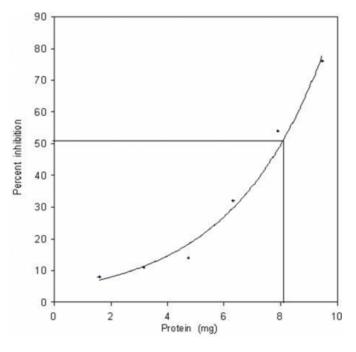
Activity of \mathbb{I}_1 -PI is measured by estimating the mg of plasma protein required to inhibit the enzyme (trypsin-20µg) to the

extent of 50% at optimum pH & optimum temperature conditions.

Antielastase activity was determined by incubating enzyme elastase and its substrate N-Succinyl -Ala-Ala-Ala-pnitroanilide with different volumes of plasma and without plasma and estimating the mg of plasma protein required to inhibit the enzyme to the extent of 50% at optimum pH and optimum temperature condition. The antielastase inhibitors of plasma inhibit the hydrolysis of N-Succinyl -Ala-Ala-Alap-nitroanilide by elastase in tris buffer, pH 8.0. The reaction is stopped by adding acetic acid, and the absorbance of pnitroaniline liberated after hydrolysis is read at 410 nm. The percent inhibition of elastase was calculated by comparing with positive control (without inhibitor) which is taken as 100% activity. The mg of protein required for 50% inhibition was found from the plot of the protein concentration (mg) against percent inhibition₁₂ (Figure 2).

Figure 2





Activity of \mathbb{I}_1 -PI is measured by estimating the mg of plasma protein required to inhibit the enzyme (elastase-1.5µg) to the extent of 50% at optimum pH and optimum temperature conditions.

Pulmonary function tests were performed in all patients and in control subjects. Forced expiratory volume in one second (FEV₁) and forced vital capacity (FVC) were measured with Ocean Win Sipro Software. FEV₁ and FVC were expressed as a percentage of the predicted values for age, sex, and height.

STATISTICAL ANALYSIS

Statistical analysis was carried out using unpaired 't' test. P value less than 0.05 (P<0.05) was considered as significant.

RESULT & DISCUSSION

Proteases and antiproteases balance is essential for the normal lung function. In the healthy lung, the proteolytic activity of proteases is balanced by presence of antiproteases locally. Under certain conditions, when there is an increased protease burden which is not compensated by antiprotease activity, lung injury follows. Also, when antiprotease defense is decreased, lung parenchymal changes may result₁₄. A number of diseases involving the lung, such as COPD, emphysema, bronchiectasis and bronchial asthma have been associated with a disturbance of these balances_{6,778}.

Human plasma is a rich source of different proteinase inhibitors (antiproteases) whose function is to control the proteolysis by affecting the proteinase activity. I_1 -PI and I_2 -M are the two important proteinase inhibitors present in plasma. I₁-PI is the chief proteinase inhibitor present in highest amount inhibiting the serine proteinases elastase and trypsin₄. I₁-PI, an "acute-phase-reactant" may increase during pregnancy, during bacterial infections, after thyphoid vaccine, in the event of severe burns, or in the presence of malignant tumors and decrease in pulmonary emphysema, cirrhosis of the liver and noninfectious hepatitis in the neonate₁₅. I₂-M is an effective inhibitor of both human neutrophil elastase and cathepsin G₅. I₂-M is not an "acutephase-reactant". Increased I2-M levels have been reported in pulmonary disorders, diabetes mellitus and in patients with liver disease₁₆.

In the present study, we estimated the plasma concentration of \mathbb{I}_1 -PI and \mathbb{I}_2 -M and also total antitryptic and total antielastase activity of plasma. Table 1 shows, in patients with COPD, emphysema, bronchiectasis and bronchial asthma, \mathbb{I}_1 -PI and \mathbb{I}_2 -M concentration is increased as compared with control. This might be the body's response to deal with increased proteolytic activity so that balance is maintained.

Figure 3

Table 1: Comparison of I-PI concentration and I-M concentration between study and control groups.

Study Groups		Control n=50	COPD n=24	Emphy- sema n=20	Bronchi- ectasis n=32	Bronchial Asthma n=35
αı-PI mg/dl	Range	130-210	130-288	132-223	146-240	133-260
	Median	165.0	196.5	196.5	191.5	184.0
	\pm SD	23.36	38.43	24.55	25.35	32.48
	p Value		p<0.001	p<0.001	p<0.001	p<0.001
∞2-M mg/dl	Range	133-270	146-330	163-313	181-373	177-400
	Median	205.0	223.5	246.5	234.5	250.0
	\pm SD	32.28	50.76	44.15	54.87	52.84
	p Value		NS	p<0.001	p<0.01	p<0.001

Values expressed as median ± SD NS= Not significant

Similar to our study Denchev and colleagues (1977) estimated the \mathbb{I}_1 -PI in patients with bronchial asthma with different forms and severity of the disease. \mathbb{I}_1 -PI was found to be elevated in all patient groups. The elevated \mathbb{I}_1 -PI concentration may be explained as a response of allergic process and secondary bacterial inflammatory reactions8.

The functional capacity of the \mathbb{I}_1 -PI is markedly susceptible to oxidative inactivation. This inactivation appears to be due to the oxidation of a critical methionine residue at the elastase inhibitory site of \mathbb{I}_1 -PI. Reduced levels of elastase inhibitory activity in the lung can result from the partial oxidation of \mathbb{I}_1 -PI through inhalation of oxidants present in tobacco smoke, ozone, or industrial gases, as well as from oxidants released by lung macrophages and other phagocytic cells₁₇.

The activity of \mathbb{I}_1 -PI and \mathbb{I}_2 -M as judged by inhibition of trypsin and elastase on their respective substrates, in vitro, was found to vary in different conditions (Table 2).

Figure 4

Table 2 : Comparison of total Antitryptic and Antielastase activity of plasma between study and control group.

Study Groups		Control n=50	COPD n=24	Emphy- sema n=20	Bronchi- ectasis n=32	Bronchial Asthma n=35
Total Anti- tryptic activity***	Range	5.4-10.5	4.4-9.5	6.25-10.6	6.35-11.3	5.7-16.9
	Median	7.475	6.00	9.125	8.3	7.9
	± SD	1.3	1.43	1.40	1.53	2.18
	p Value		p<0.001	p<0.001	p<0.001	NS
Total Anti- elastase Activity*	Range	4.8-11.55	5.0-15.0	6.1-12.05	5.45-13.05	5.15-12.9
	Median	7.8	6.75	9.6	7.9	8.6
	± SD	1.53	2.38	1.84	1.70	1.75
	p Value		p<0.001	p<0.001	NS	p<0.001

Values expressed as median ± SD

NS=Not significant

**: mg of plasma protein required to inhibit the enzyme (trypsin - 20µg, elastase - 1.5µg) to the extent of 50% at optimum pH & optimum temperature conditions.

In the condition of COPD, both antitryptic and antielastase activity of plasma increased significantly (p<0.001) as compared to control, which may be because of increased concentrations of both \mathbb{I}_1 -PI and \mathbb{I}_2 -M. But in the other groups antitryptic activity decreased inspite of increased concentration of the inhibitors. Antielastase activity was not found to vary significantly in conditions of bronchiectasis and was found to decrease significantly (p<0.001) in emphysema and bronchial asthma. Similar to our study Gaillard and colleagues showed that plasma elastase inhibitory capacity was markedly reduced inspite of increased with nonasthmatic individuals₁₈.

Literature shows smoking is associated with the loss of lung elasticity in \mathbb{I}_1 -antitrypsin deficiency₁₉ and also to the decrease in lung function. Pulmonary function tests are useful in patients with chronic airway obstruction to confirm the obstructive abnormality, to quantify the severity of the defect, to assess the reversibility of the airflow obstruction in response to therapy₂₀, and to monitor the course of the disease.

In the present study we estimated pulmonary function parameters of patients and control group (Table 3).

Figure 5

Table 3 : Pulmonary Function Tests in different study groups.

Study Groups		Control n=50	COPD n=24	Emphy- sema n=20	Bronchi- ectasis n=32	Bronchial Asthma n=35
FEV1	Range	91.0-101.0	28.0-80.0	11.0-65.0	15.0-108.0	21.0-93.0
	Median	96.00	48.50	29.50	67.00	59.00
	\pm SD	2.22	16.37	17.58	21.90	17.29
	p Value		p<0.001	p<0.001	p<0.001	p<0.001
FVC	Range	88.0-99.0	42.0-95.0	28.0-85.0	13.0-106.0	29.0-94.0
	Median	97.00	61.00	47.00	77.50	70.00
	± SD	2.83	16.16	16.90	20.96	16.77
	P Value		p<0.001	p<0.001	p<0.001	p<0.001

And we also investigated whether antiproteases concentration and antiproteases activity are related to lung function tests in all groups (Table 4).

Table 4 : Correlation coefficient (R) between different parameters and PFT (% Predicted) in different study groups.

Pulmonary function tests of the study group shows significant reduction in FEV_1 and FVC as compared to control group. This clearly indicates obstruction of airways in all the conditions under study i.e. COPD, emphysema, bronchiectasis and bronchial asthma. The cause and mechanism of development of these pathologies may be different. Correlation coefficient values (Table 4) showed that significant correlation between aniproteases concentration, antiproteases activity and pulmonary function tests was found only between some parameters in different study groups.

Thus uniformity is seen in all the disorders under study that there is blockage of airways and there is shift in proteasesantiproteases balance towards proteases as reflected by the measurement of antitryptic and antielastase activity. Whether this shift is cause or effect of the disorder remains to be studied.

CORRESPONDENCE TO

Madhavi S. Phadke, PhD 727, Dhanwantari Society, 'B' wing, Sector II, Plot 3&4, Deonar Municipal Colony, Mumbai - 400043. India E-mail:

mangalphadke@hotmail.com 91-22-56049569,

91-22-25559278 **References**

1. Gadek J E & Pacht E R (1996) Chest 110, 248S-250S

2. Cynthia Llyod & James Travis (1989) Clin Chem 35, 1971-1975

3. Buhl R, Meyer A & Volgelmeier C (1996) Chest 110, 267S-272S

4. Robert K M, Daryl K G, Peter A, Mayes, Victor M R (2000) Harper's

Biochemistry, 25th edn, pp. 744-745, Appleton and Lange, Stamford,

Connecticul.

5. James T (1988) The American Journal of Medicine 84 (Suppl 6A), 37-42

6. Gadek J E & Pacht E R (1990) Lung 168(Suppl.), 30-42

7. Kilore-Smith T A, Dowdeswell R J & Gaillard M C (1989) Clinica Chimica

Acta 185(1), 81-90

8. Denchev K, Radkov & Lipcheva N (1977) Vutreshni Bolesti 16(2), 75-78

9. Global Initiative for chronic obstruction lung disease (COLD) WHO/NHLBI

Initiative 2001. Current Medical Scene, Apr-June 2002, 17(2):4

10. Cortran, Kumar & Robbins (1989) Robbins pathological basis of disease, 4th

edn, pp. 755-810, W.B.Saunders Company, Philadelphia 11. Ross & Wilson (2001) Anatomy and physiology in health and illness, 9th edn,

pp. 240-268, Churchil Livingston

12. Tarvis J & Johnson D (1981) Methods in enzymology 80 , 755-757

13. Albert A. Dietz, Herbert M, Rubinstein L A & Verue Hodges (1974) Clin

Chem 20(3), 396-399

14. Bhul R, Meyer A & Volgelmeier C (1996) Chest 110(Suppl), 267S-272S

15. Jacobo W, Lois Y M & Friedrich K (1991) Clin Chem 37(1) 108-110

16. Harpel P C (1976) Methods Enzymol 45, 639-652

17. Theron A & Anderson R (1985) Am Rev Respir Dis 132(5), 1049-1054

18. Gaillard M C, Kilore-Smith T A, Nogueira C, Dunn D, Jenkins T, Fine B &

Kllenbach J (1992) Am Rev Respir Dis 145(6), 1311-1315 19. Salahuddin P (1991) Indian J. Biochem Biophys 88, 164-167

20. Burge P S, Calverley P M, Jones P W, et al. (2000) BMJ 320, 1297

Author Information

Raghunath R. Rai, M.Sc. Research Scholar, Department of Biochemistry, Lokmanya Tilak Municipal Medical College

Madhavi S. Phadke, Ph.D.

Associate Professor, Department of Biochemistry, Lokmanya Tilak Municipal Medical College