

Prevalence of *Campylobacter* spp. in Nigerian Indigenous Chicken in Sokoto State Northwestern Nigeria.

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Citation

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Abstract

The study was carried out to determine the prevalence of *Campylobacter* spp. in Nigerian indigenous chickens and to characterize the isolated strains using phenotypic methods and biotyping. Of the 866 samples collected 672(77.6%) were campylobacter positive. A total of 828 strains of *Campylobacter* spp were isolated which were identified using biochemical methods. The species identified from the study were *C. jejuni* 556(67.2%); *C. coli* 179(21.6%); *C. lari* 62(7.5%) and *C. upsaliensis* 31(3.7%). The biotyping of isolates yielded *C. jejuni* (biotype I, 355(63.9%); biotype II, 139(25.0%); biotype III, 54(9.7%) and 8(1.4%) for biotype IV); *C. coli* (biotype I, 102(57.0%) and biotype II, 77(43.0 %)) and *C. lari* (biotype I, 37 (59.7%) and biotype II, 25(40.3%)). The study has demonstrated the carriage of *Campylobacter* in Nigerian indigenous chicken. The role of the Nigeria indigenous chicken in the transmission of the campylobacter is unknown, but the frequency with which birds are associated with these organisms suggests that they may have an important role in their dissemination.

INTRODUCTION

Campylobacter spp are recognized worldwide as the major cause of human enteritis (Hascelik et al. 1991: Pearson and Healing, 1992: Taylor, 1992: Logan et al.1999). Although several animal species have been shown to carry campylobacters and a variety of vehicles of human infection have been demonstrated (Garcia et al. 1983: Atabay and Corry, 1997, 1998; Ridsdale et al. 1998, 1999). Avian carriage of *Campylobacter* has been regarded as a potential hazard to human health, either through consumption of undercooked carcass or by contamination of water supplies (Skirrow, 1994: Varslot et al. 1996). A wide variety of avian species, including domestic chickens, turkeys, ducks, pigeons, quail, waterfowls, geese and ostriches, harbour *Campylobacter* spp (Pacha et al. 1988: Yogasundram et al.1989: oyarzabal et al. 1995; Aydin et al. 2001: Broman et al. 2004). However, they are unevenly distributed among species, and the feeding behaviour of birds has been shown to influence the *Campylobacter* colonization rate (Waldenstrom et al. 2002).

The Nigerian indigenous chickens are raised on a small in most households in rural and semi-urban areas of Northwestern Nigeria. The chickens are reared as free range

and may therefore; contaminate water and the surrounding environment. Their habits bring them into close contact with human, grazing animals and even dogs and cats. They may constitute a potential public health risk in relation to *Campylobacter* infections in humans and animals. The study was therefore conducted to determine the prevalence of thermophilic *Campylobacter* and their biotypes in Nigeria indigenous chicken.

MATERIALS AND METHODS

Between December, 2007 and November, 2008, 866 indigenous chickens were sampled across the state for *Campylobacter* spp. Faecal material was obtained from the chicken by cloacal swab. The faecal material so obtained was placed directly into Amies transport medium (Oxoid, CM425) and transported to the laboratory immediately. At the laboratory the transport broth were incubated at 37oC for 72h before subculture to mCCDA (modified charcoal cefoperazone deoxycholate agar; Oxoid, CM739 plus SR155) and incubated at 42oC for 72h microaerobically (CampyGen; oxoid; CN35A) in an anaerobic jar, and were examined after 24, 48 and 72h incubation. Suspect colonies were Gram stained and tested for the production of oxidase and catalase. Colonies giving reaction typical for

Campylobacter were purified by streaking onto blood agar. All the isolates were characterized using standard *Campylobacter* phenotypic identification procedures described by Atabay and Corry (1997), such as hippurate hydrolysis, rapid production of hydrogen sulphide DNA hydrolysis, aerobic growth at 37°C, microaerobic growth at 37°C and 43°C.

Biotyping of isolates was carried out using the extending biotyping scheme of Lior, (1984). The scheme is based on hippurate hydrolysis; rapid production of hydrogen sulphide in FBP broth and DNA hydrolysis.

RESULTS

Of the 866 chickens examined, 672 (77.6%) were found to be carrying campylobacters. More than one species of *Campylobacter* were isolated from 260 (30%) of the samples. The species of *Campylobacter* species identified in this study are *C. jejuni*, *C. coli*, *C. lari*, and *C. upsaliensis*. A total of 828 strains of *Campylobacter* spp were isolated which were identified using biochemical methods. The isolation rates of the *Campylobacter* spp are *C. jejuni* 556(67.2%); *C. coli* 179(21.6%); *C. lari* 62(7.5%) and *C. upsaliensis* 31(3.7%) as shown in table I.

Figure 1

Table I: Isolation rates of spp from Nigerian indigenous chickens.

<i>Campylobacter</i> spp.	Number and percentage isolates
<i>Campylobacter jejuni</i>	556(67.2%)
<i>Campylobacter coli</i>	179(21.6%)
<i>Campylobacter lari</i>	62(7.5%)
<i>Campylobacter upsaliensis</i>	31(3.7)
Total	828(100%)

Bioyping of the isolates were carried out for *Campylobacter jejuni*, *Campylobacter coli*, and *Campylobacter lari* strains. The *C. jejuni* biotypes were biotype, I 355(63.9%); biotype II, 139(25.0%); biotype III, 54(9.7%) and 8(1.4%) for biotype IV (Table II). The *C. coli* biotypes were biotype I, 102(57.0%) and biotype II, 77(43.0 %); the biotypes of *C. lari* were 37 (59.7%) and 25(40.3%) for biotype I and II respectively (Table II).

Figure 2

Table II: Percentage distribution of

<i>Campylobacter</i> strains	Biotypes	Isolation rate (%)
<i>Campylobacter jejuni</i>	I	355(63.9%)
	II	139(25.0%)
	III	54(9.2%)
	IV	8(1.4%)
<i>Campylobacter coli</i>	I	102(57.0%)
	II	77(43.0%)
<i>Campylobacter lari</i>	I	37(59.7%)
	II	25(40.3%)

DISCUSSION

The prevalence of *Campylobacter* spp in Nigerian indigenous chicken from this study is 77.6%, thus demonstrating the significance of the chickens as reservoirs of *Campylobacter* spp. The carrier rate of *Campylobacter* organism examined in this study is higher than those obtained from broiler, 94.2% (workman et al. 2005), but lower than 45.9% by Atanassova and Ring (1999) and 60% by Georgios (2004) from broiler chickens. The free-range nature of rearing the chickens exposes them to both human and animal wastes and other potential sources of enteric pathogens. The management system can lead to the contamination of environmental sources such as water by the faeces of the chickens and the organism can easily be transmitted to humans and animals via this environmental sources. Varslot et al., (1996), described water-borne outbreaks of *Campylobacter jejuni* infections in humans in Norway. These epidemics were traced to contamination of drinking water by faeces of birds (geese). In Nigeria, the indigenous chickens are always in close contact with other animals and humans, and considering the zoonotic nature of *Campylobacter*, It can be contracted through close contacted with infected animals (Blaser et al., 1983).

The high isolation rate of 67.2% of *C. jejuni* from chickens in this study is in agreement with reports of other studies by Nielsen et al. (1997), Shane, (2000), Wedderkopp et al. (2001), Aydin et al. (2001) and Geogios et al.(2004). The low isolation rate of *C. coli* from chickens in this study is however, in line with the findings that *C. coli* has a lower rates of isolation than *C. jejuni*, from chicken as reported by Atanassova and Ring, (1999), Christopher et al., (1982) and Smith, (1995). The very low isolation rate of *C. upsaliensis* (3.0%) from chickens in this study was expected, because the body temperature of chicken does not favour their survival in chicken. The free range nature of local chicken may possibly expose the chicken to *C. upsaliensis* through feeding on faeces of other domestic animals and even pets which are reported to harbour *C. upsaliensis* (Workman et al., 2005). The observations in this study agreed with

observations of Atanassova and Ring, (1999) and Baserisalihe et al., (2007) who also reported very low isolation rates of *C. upsaliensis* from poultry birds.

The biotyping scheme of Lior (1984), divided the *C. jejuni* from this study into Biotypes I, II, III, IV and *C. coli* into two biotypes I and II. This observation is an indication that the indigenous chicken can harbour diverse strains of *Campylobacter*. Lior (1984), observed that *Campylobacter jejuni* strains isolated from humans and animals were biotype I. The isolation of *Campylobacter jejuni* biotype I from the indigenous chickens is of serious public health importance, as *Campylobacter jejuni* biotype I has been implicated in human campylobacteriosis. The indigenous chicken should be regarded as a potential reservoir for human and animal infection with *Campylobacter*. *Campylobacter jejuni* is the main causative agent of food-borne gastroenteritis in humans, and also causes a variety of disease, such as enteritis, abortion, septicaemia and mastitis, in animals (Aydin, et al, 2001).

The role of the Nigeria indigenous chicken in the transmission of the *Campylobacter* is unknown, but the frequency with which birds are associated with these organisms suggests that they may have an important role in their dissemination. This study has demonstrated the carriage of *Campylobacter* in Nigerian indigenous chicken which have not previously been investigated.

References

- r-0. Atabay, H. L., and Corry, J. E. L. (1997). The prevalence of *Campylobacters* and *arcobacters* in broiler chickens. *Journal of Applied Microbiology*. 83, 619-626.
- r-1. Atabay, H. L., and Corry, J. E. L. (1998). The isolation and prevalence of *campylobacters* from the dairy using a variety of methods. *Journal of Applied Microbiology*. 84,733-740.
- r-2. Atanassova, V. and Ring, C. (1999): Prevalence of *Campylobacter* spp. in poultry meat in Germany. *International Journal of Food Microbiology*. 51, 187-190
- r-3. Aydin, F., Atabay, H.I. and Akan, M. (2001). The isolation and characterization of *C. jejuni* subsp. *jejuni* from domestic geese (*Anser anser*). *Journal of Applied Microbiology*. 90, 637-642
- r-4. Baserisalehi, M., Bahador, N. and Kapadnis, B.P. (2007) Isolation and characterization of *Campylobacter* spp. from domestic animals and poultry in south of Iran. *Pakistan Journal of Bioogical Science*. 10(9), 1519-1524.
- r-5. Blaser, M.j., Taylor, D.N. and Feldman, R.A. (1983) Epidemiology of *Campylobacter jejuni* infections. *Epidemiologic reviews*. 5, 157-176
- r-6. Broman, T., Weldenstrom. J., Dahlgren, D. et al (2004) Diversities and similarities of PFGE profiles of *Campylobacter jejuni* isolated from migrating birds and humans. *Journal of Applird Microbiology*. 96, 834-843.
- r-7. Christopher, F. M., Smith, G. C., Vanderzant, C. (1982) Examination of poultry giblets, raw milk and raw meat for *Campylobacter fetus* subsp. *jejuni*. *Journal of Food protection*. 45, 260-262.
- r-8. Georgios, K., Dang, D.B., Mariannr, L., Mogens, M., Henrick, B., Pieter, T. and Claus, B.V.C. (2004) Use of PCR analysis, and DNA microarrays for detection of *Campylobacter jejuni* and *Campylobacter coli* from Chicken feces. *Journal of Clinical Microbiology*. 42(9), 3985-3991.
- r-9. Hascelik, G., Akyon, Y., Hayran, M., Yurdakok, K. and Berkman, E. 1991) *Campylobacter* as acause of acute enteritis in turkey. *Therapy of Infectious Diseases*. 6, 288-292.
- r-10. Lior, H.(1984) Neqw extended Biotyping scheme for *Campylobacter jejuni*, *C. coli*, and *C. lari*. *Journal of Clinical Microbiology*. 20, 636-640
- r-11. Logan, J.M.J., Lawson, A.J., O'Neill, G.L. and Stanley, J. (1999) A large scale survey of *Campylobacter* in human gastroenteritis using PCR for screening and PCR-ELISA for identification. In Abstracts of the 10th International Workshop on CHRO, Baltimore, Maryland, US. CD19, 28
- r-12. Oyarzabal, O.A., Conner, D.E. and Hoerr, F.J. (1995) Incidence of *campylobacters* in the intestine of avian species in Alabama. *Avian Diseases*. 39, 147-151.
- r-13. Pacha, R.E., Clark, G.W., Williams, E.A. and Carter, A.M. (1988) Migratory birds of cental Washimington as reservoirs of *Campylobacter jejuni*. *Canadian Journal of Microbiology*. 34, 80-82.
- r-14. Pearson, A.D. and Healing, T.D. (1992) The surveillance and control of *Campylobacter* infection. *CDR Review: Communicable Diseases* report, UK 2, Revuew no. 12.
- r-15. Ridsdale, J.A., Atabay, H.I. and Corry, J.E.L. (1998) Prevalence of *campylobacters* and *arcobacters* in ducks at the abattoir. *Journal of Applied Microbiology*. 85, 567-573.
- r-16. Ridsdale, J.A., Atabay, H.I. and Corry, J.E.L. (1999) *Campylobacter* and *Arcobacter* spp. Isolated from the carcasses and caeca of commercially reared ducks (Poster Presentation). *Anaerobe* 5, 317-320
- r-17. Shane, S. M., (2000): *Campylobacter* infection of commercial poultry. *Science Review, Technical Office of International Epizootic*.

19, 376-395

r-18. Skirrow, M.B. (1994) Diseases due to campylobacter, *Helicobacter* and related bacteria. *Journal of Comparative Pathology*. 2, 9- 11.

r-19. Smith, J. L. (1995) Arthritis, Guillain-Barré Syndrome, and other sequelae of *Campylobacter jejuni* enteritis. *Journal of Food Protection*. 58(10), 1153-1170.

r-20. Taylor, D.N. (1992) *Campylobacter* infection in developing countries. In: Nachamkin, I and Blaser, M.J. (eds) *Campylobacter jejuni: Current status and future trends* Washington: American Society for Microbiology. Pp 20-30.

r-21. Varslot, M., Resell, J. and Fostad, F.G. (1996) Water-borne outbreaks of

Campylobacter gastroenteritis due to pink-footed geese in Norway in 1994 and 1995. *Tidsskrift for Den Norske Lægeforening*. 116, 3366-3369.

r-22. Waldenstrom, J, Broman, T., Carlsson, I. et al, (2002)

Prevalence of *C.jejuni*, *C.*

lari, and *C. coli* in different ecological guilds and taxa of migrating birds dagger. *Applied Environmental Microbiology*. 68, 5917-5917.

r-23. Wedderkopp, A., Gradel, K. O., Jorgensen, J. C., Madsen, M. (2001) Pre-harvest surveillance of *Campylobacter* and *Salmonella* in Danish broiler flocks: a 2-year study. *International Journal of Food Microbiology*. 68, 53-59.

r-24. Workman, N.S., Mathison, E.G., and Lavoie, C.M. (2005) Pet dogs and chicken meat as reservoir of *Campylobacter* spp. in Barbados. *Journal*

of *Clinical Microbiology*. 43(6), 2642-2650.

r-25. Yogasundram, K., Shane, S.M. and Harrington, K.S. (1989) Prevalence of *Campylobacter jejuni* in selected domestic and wild birds in Louisiana. *Avian Diseases*. 33,664-667

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