Surgical Site Infection Complicating Leech Therapy

J Orsini, G Sakoulas

Citation

J Orsini, G Sakoulas. *Surgical Site Infection Complicating Leech Therapy*. The Internet Journal of Plastic Surgery. 2006 Volume 3 Number 1.

Abstract

Aeromonas hydrophila infections are a recognized complication of postoperative leech application, and can occur with measurable frequency in populations of patients treated with leeches. We report a case of an abdominal flap infection caused by multiple organisms, including A. hydrophila, in a patient treated with leech therapy. Prompt surgical evaluation of wounds in combination with appropriate antibiotic therapy is recommended for the management of these infections.

CASE REPORT

A 57-year-old man was admitted for an elective abdominal wall reconstruction of a large ventral incisional hernia, which complicated emergent surgical repair of an abdominal aortic aneurysm rupture 2 years prior admission. One day after admission, he underwent a free tensor fascia lata (TFL) flap reconstruction. On hospital day 4th, he underwent exploratory laparotomy due to abdominal distention and superficial necrosis of the flap, with exploration and debridement of the TFL flap secondary to venous congestion. Leech therapy was initiated to alleviate the venous congestion of the TFL flap.

The postoperative course was complicated by persistent fevers (103°F), leukocytosis, worsening renal function, and purulent discharge from the surgical site. Intravenous vancomycin (1 g Q24h) and piperacillin/tazobactam (4.5 g Q8h) were initiated empirically. Relevant laboratory data included a white blood cell count of 12,4000/mm³, a hemoglobin of 13.8 g/dl, a platelet count 190,000/mm³, a serum creatinine 2.4 mg/dl, a creatine kinase 1118 units/L, and a C-reactive protein of 13.0 mg/dl. Liver function and coagulation studies were within normal limits. Chest radiography was normal.

His clinical condition improved after several days, with resolution of fever and renal function. Abdominal wound culture grew Aeromonas hydrophila, Pseudomonas aeruginosa, Proteus mirabilis and gamma-hemolytic streptococcus. Abdominal flap culture demonstrated A. hydrophila and P. mirabilis. Blood and urine cultures were negative. The patient was discharged on intravenous antibiotic regimen composed of meropenem (1 g Q8h) and ciprofloxacin (400 mg Q12h) for 3 weeks, followed by ciprofloxacin (750 mg PO Q12h) for an additional 4 weeks. After several weeks, the patient was electively admitted for a removal of the infected mesh with a placement of an alloderm graft.

DISCUSSION

Aeromonas spp. are gram-negative, motile, facultative anaerobic, oxidase positive rods associated with a wide variety of human infections, often in conjunction with other organisms ($_{1, 2}$). They are usually considered to be opportunistic pathogens but infections in immunologically competent individuals have been described ($_{1, 3, 4}$). They are ubiquitous organisms isolated worldwide from aquatic environments and soil, including hospital water supplies, especially during warmer months ($_{5, 6, 7, 8}$). Currently 14 species have been named, but only A. hydrophila, A. caviae and A.veronii are of major clinical significance to humans ($_{9}$). Aeromonas spp. posses several virulence factors such as enterotoxins, aerolysin, hemolysin and mucinase, which mediate the pathogenesis of human infections ($_{10}$).

Gastroenteritis is the most common clinical manifestation of Aeromonas infection, resembling traveler's diarrhea and hemolytic uremic syndrome ($_7$, $_{11}$, $_{12}$). Diarrhea is usually watery and self-limited but patients may developed abdominal pain and bloody stools ($_8$). Bacteremia ($_9$), urinary tract infection ($_{13}$), meningitis ($_{14}$), hepatobiliary system infection ($_{15}$), peritonitis ($_{16}$), endocarditis ($_{17}$), septic arthritis ($_{18}$) and respiratory infections ($_9$) have also been reported. Soft tissue infections from Aeromonas spp. are sporadic and infrequent, but typically occur as a result of contamination of broken skin by water containing the organism. These infections range in presentation from cellulitis mimicking streptococcal or staphylococcal soft-tissue infections (19) to more severe infections such as myonecrosis, fasciitis and ecthyma gangrenosum. The more severe infections have been described in immunosupressed patients (2, 8, 20). Wound infections mostly affect lower extremities (21) and often progress rapidly requiring surgical debridement or amputation of the involved limb (22). They have been described among tsunami survivors (23) and patients in the burn units (24).

Medicinal leeches have an important and expanding role in medicine, but infection can complicate their use. Leeches have been used by surgeons, especially those in plastics and reconstruction, to decrease venous congestion of skin flaps and to improve micro-revascularization of flaps, grafts and replants. Hirudin, a powerful anticoagulant that inhibits thrombin, is secreted in salivary secretions of leeches and injected into the flap. Hemoglobin that is sucked by the leeches is denaturated by Aeromonas. Of the hemoglobin degradation products, heme is utilized by Aeromonas and the protein components of globulin, is utilized by the leech $(_{25}, _{26})$. Each leech directly extracts 5-10 ml of blood from the flap, and an extra 20-50 ml of blood is lost from oozing from the bite site after the leech detaches. The leech saliva also contains collagenase and hyaluronidase, which facilitate local infiltration of antithrombotic mediators into the congested tissue $(_{27}, _{28}, _{29})$.

As this case illustrates, patients receiving medicinal leech therapy are at risk of developing Aeromonas infections, and up to 20% of patients treated with medicinal leeches may develop gram-negative bacterial infections ($_{30}$). This is reflective of the fact that Aeromonas spp are a normal inhabitants of the foregut of leeches ($_8$). Infections complicating leech therapy can range from minor wound complications to extensive tissue loss and sepsis, especially in patients with compromised arterial blood supply to the affected area ($_{30}$). The onset can range from 24 hours to over 10 days after leech application. Late infections may represent bacterial invasion from colonized necrotic tissue ($_{27}$, $_{31}$, $_{32}$). Prophylactic antibiotics such as cefotaxime, third generation cephalosporins and ciprofloxacin have been recommended at the time of leech application ($_{32}$).

Antimicrobial therapy of Aeromonas infections may be

difficult, especially empirically, because of intrinsic resistance to penicillin and other beta-lactams as a result of a chromosomal \mathbb{I} -lactamase (3, 33, 34). Furthermore, the use of \mathbb{I} lactamase inhibitors plays no role in inhibiting the Aeromonas -lactamase and therefore rarely enhances -lactam antibiotic activity (35). Antibiotics such as ampicillinsulbactam, piperacillin or cefotetan, which are often administered in the empiric treatment of intrabdominal infections, have reduced activity against Aeromonas.

Most Aeromonas are susceptible to quinolones, aztreonam, carbapenems, third generation cephalosporins, tetracyclines, chloramphenicol and aminoglycosides ($_{35}$, $_{36}$, $_{37}$). It is important to emphasize the necessity of prophylactic antibiotic therapy at the time of leech therapy using agents with activity against Aeromonas in order to decrease the likelihood of serious complications associated with this treatment.

CORRESPONDENCE TO

George Sakoulas, MD Division of Infectious Diseases New York Medical College Munger 245 Valhalla, NY 10595 Phone: 914-493-8865 Fax: 914-594-4973 Email: george_sakoulas@nymc.edu

References

1. Joseph SW, Daily OP, Hunt WS, Seidler RJ, Allen DA, Colwell RR. Aeromonas Primary Wound Infection of a Diver in Polluted Waters. J Clin Microbiol. 1979;10:46-9 2. Gold WL, Salit IE. Aeromonas hydrophila infections of the skin and soft tissue: report of 11 cases and review. Clin Infect Dis. 1993;16:69-74

3. Bulger RJ, Sherris JC. The clinical significance of Aeromonas hydrophila. Report of two cases. Arch Intern Med. 1966;118:562-4

4. McCracken AW, Barkley P. Isolation of aeromonas species from clinical sources. J Clin Pathol. 1972;25:970-5
5. Hazen TC, Flierman CB, Hirsch RP, Esch GW. Prevalence and distribution of Aeromonas hydrophila in the United States. Appl Environ Microbiol. 1978;36:731-8
6. Van der Kooj D. Properties of Aeromonads and their occurrence and hygienic significance in drinking water. Zentralb Bakt Hyg B. 1988;187:1-17

7. Yadav A, Kumar A. Prevalence of enterotoxigenic motile Aeromonas in children, fish, milk and ice cream and their public health significance. Southeast Asian J Trop Med Public Health. 2000;31:153-6

8. Steinberg JP, Del Rio C. Other Gram-Negative and Gram-Variable Bacilli. In: Mandell G, Bennett J, Dolin R et al. Principles and Practice of Infectious Diseases. Philadelphia: Churchill-Livingstone; 2005. p2751-63

9. Janda JM, Abbott SL. Evolving concepts regarding the genus Aeromonas: an expanding Panorama of species, disease presentations, and unanswered questions. Clin Infect Dis. 1998;27:332-44

10. Rabaan A, Gryllas I, Tomas T, Shaw J. Motility and the polar flagellum are required for Aeromonas caviae adherence to HEP-2 cells. Infection and Immunity.

2001;4257-67

11. Yamada S, Matsushita S, Dejsirilert S, Kudoh Y. Incidence and clinical symptoms of Aeromonas-associated Traveller's diarrhoea in Tokyo. Epidemiol Infect. 1997;119:121-6

12. Bogdanovic R, Cobeljic M, Markovic M, Nikolic V, Ognjanovic M, Sarjanovic L, et al. Haemolytic-uraemic syndrome associated with Aeromonas hydrophila

enterocolitis. Pediatr Nephrol. 1991;5:293-5

13. Hua HT, Bollet C,, Tercian S, Drancourt M, Raoult D. Aeromonas popoffii urinary tract infection. J Clin Microbiol. 2004;42:5427-8

14. Parras F, Diaz MD, Reina J, Moreno S, Guerrero C, Bouza E. Meningitis due to Aeromonas species: case report and review. Clin Infect Dis. 1993;17:1058-60

15. Clark N, Chenoweth CE. Aeromonas Infection of the Hepatobiliary System: Report of 15 Cases and Review of the Literature. Clin Infect Dis. 2003;37:506-13

16. Munoz P, Fernandez-Baca V, Pelaez T, Sanchez R, Rodriguez-Creixems M, Bouza E. Aeromonas peritonitis. Clin Infect Dis. 1994;18:32-7

17. Ong KR, Sordillo E, Frankel E. Unusual case of Aeromonas hydrophila endocarditis. J Clin Microbiol. 1991;29:1056-7

18. Elwitigala JP, Higgs DS, Namnyak S, White JW, Yaneza A. Septic arthritis due to Aeromonas hydrophila: case report and review of the literature. International Journal of Clinical Practice. 2005;59:121

19. Vally H, Whittle A, Cameron S, Dowse G, Watson T. Outbreak of Aeromonas hydrophila Wound Infections Associated with Mud Football. Clin Infect Dis. 2004;38:1084-9

20. Musher DM. Cutaneous and soft-tissue manifestations of sepsis due to gram-negative enteric bacilli. Rev Infect Dis. 1980;2:854-66

21. Khardori N, Fainstein V. Aeromonas and Plesiomonas as etiological agents. Ann Rev Microbiol. 1988;42:395-419 22. Isaacs RD, Paviour SD, Bunker DE, Lang SD. Wound infection with aerogenic Aeromonas strains: a review of twenty-seven cases. Eur J Clin Microbiol Infect Dis. 1988:7:355-60

23. Lim PL. Wound Infections in Tsunami Survivors. A Commentary. Ann Acad Med Singapore. 2005;34:582-5 24. Barillo DJ, McManus AT, Cioffi WG, McManus WF,

Kim SH, Pruitt BA Jr. Aeromonas bacteraemia in burn patients. Burns. 1996;22:48-52 25. Richerson JT, Davis JA, Meystrik R. Aeromonas,

25. Richerson JT, Davis JA, Meystrik R. Aeromonas, acclimation, and penicillin as complications when leeches are apply to skin flaps in rabbits. Lab Anim. 1990;24:147-50
26. Markwardt F. Hirudin as an inhibitor of thrombin. Methods Enzymol. 1978;19:924-32

27. Derganc M, Zdravic F. Venous congestion of flaps treated by application of leeches. Br J Plastic Surg. 1960;13:187-92

28. Utley DS, Koch RJ, Goode RL. The failing flap in facial and reconstructive surgery: Role of Medicinal Leech. Laryngoscope. 1998;108:1129-35

29. Rigbi M, Orevi M, Eldor A. Platelet aggregation and coagulation inhibitors in leech saliva and their roles in leech therapy. Semin Thromb Hemost. 1996;22:273-8
30. Mercer NS, Beere DM, Bornemisza AJ, Thomas P. Medicinal leeches as sources of wound infections. BMJ. 1987;294:937-8

31. Snower DP, Ruef C, Kuritza AP, Edberg SC. Aeromonas hydrophila infection Associated with the Use of Medicinal Leeches. J Clin Microbiol. 1989;27:1421-2

32. Lineaweaver WC, Hill MK, Buncke GM, Follansbee S, Buncke HJ, Wong RK, et al. Aeromonas hydrophila infections following use of medicinal leeches in replantation and flap surgery. Ann Plast Surg. 1992;29:238-44

33. Jones BL, Wilcox MH. Aeromonas infections and their treatment. J Antimicrob Chemother. 1995;35:453-61

 Janda JM, Guthertz LS, Kokka RP, Shimada T. Aeromonas species in septicemia: laboratory characteristics and clinical observations. Clin Infect Dis. 1994;19:77-83
 Vila J, Marco F, Soler L, Chacon M, Figueras MJ. In vitro antimicrobial susceptibility of clinical isolates of

Aeromonas caviae, Aeromonas hydrophila and Aeromonas veronii biotype sobria. J Antimicrob Chemother. 2002;49:701-2

36. Mani S, Sadigh M, Andriole VT. Clinical spectrum of Aeromonas hydrophila infections: Report of 11 cases in a community hospital and review. Infect Dis Clin Pract. 1995;4:79-86

37. Koehler JM, Ashdown LR. In vitro susceptibilities of tropical strains of Aeromonas species from Queensland, Australia, to 22 antimicrobial agents. Antimicrob Agents Chemother. 1993;37:905-7

Author Information

Jose Orsini, M.D.

Division of Infectious Diseases, New York Medical College and Westchester Medical Center

George Sakoulas, M.D.

Division of Infectious Diseases, New York Medical College and Westchester Medical Center