# Antimicrobial Susceptibility Patterns Of Bacteria To Seed Extracts Of Ricinus Communis: Findings Of A Preliminary Study In Nigeria

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#### Citation

G Jombo, M Enenebeaku. Antimicrobial Susceptibility Patterns Of Bacteria To Seed Extracts Of Ricinus Communis: Findings Of A Preliminary Study In Nigeria. The Internet Journal of Microbiology. 2006 Volume 4 Number 1.

#### Abstract

Aim: To ascertain the antibacterial properties inherent in seed extracts of Ricinus communis.

Procedure: Dry seeds of R. communis were deshelled, grounded to powder and extracted both with alcohol and water using Soxhlet machine. Different concentrations of the extracts were tested against selected bacteria using diffusion method of susceptibility testing on sensitivity testing agar medium.

#### Results:

Klebsiella pneumoniae, Escherichia coli, Proteus mirabilis, Staphylococcus aureus and Enterococcus faecalis were highly susceptible to both the methanol and water extracts of the seed while Pseudomonas aeruginosa showed reduced susceptibility.

Conclusion: The active antimicrobial ingredients in R. communis should be identified while its medicinal value to humans properly investigated in this regard.

# INTRODUCTION

Ricinus communis popularly called Castor bean in English is a plant that is widely distributed in virtually in all continents of the world<sub>1</sub>. Its leaves are verticulate, long- pertiolate with palmately divided laminae; flowers in terminal panicles are usually overtopped by lateral shoots; male flowers clustered at the bottom, female flowers above, with inconspicuous caduceus perianth. The fruits are globosse, triocular, softspiny capsules; the seeds are ovoid, compressed dorsally, thick, shiny pale-grey to almost black with dark mottling<sub>2,3,4,5</sub>.

R.communis has found several daily applications in human activity<sub>677</sub>: the oil seed is used in coating fabrics and other protective coverings<sub>8</sub>; the hydrogenated oil is utilized in the manufacture of waxes, polishes, carbon paper, candles and crayons<sub>9,10,11</sub>. It has been used with belief to cure several ailments: arthritis, asthma, boils burns, cancer, carbuncles, catarrh, chancre, cholera, cold, colic, convulsions, and 'craw-craw', to mention but a few<sub>12,13,14</sub>.

Several biophysical properties have been associated with R.

communis. In Jos, Nigeria, whole seeds in stat doses were found to stop pregnancy for about a year in two separate studies among volunteers<sub>15,16</sub>. In Norway<sub>17</sub>, it was found that the B chain of R. communis activates human complement thus boosting immunity; while findings from Sudan<sub>18</sub> revealed that, R. communis from various plant extracts were found so attain selectively various portions of the cell bodies of hippocampal neurons with defined functions. Also in India<sub>20</sub>, its anti-inflammatory and free radical scavenging activity by inhibition of lipid peroxidation was well demonstrated.

From the microbiological stand point, there has not been much published data on R. communis  $_{21,22,23}$  especially as concerns the antimicrobial properties of its various extracts on bacteria, fungi, viruses and parasites.

In view of the fact that bacteria have assumed an unprecedented level of antimicrobial resistance more than ever in the history of modern medicine<sub>24,25,26,27</sub>; the continuous search for more reliable antibiotics becomes a worthwhile and noble mission. This study was therefore set up to ascertain the antibacterial properties of the seed extracts of R. communis.

# MATERIALS AND METHOD

Setting The study was carried out in Jos Plateau state of Nigeria between August and November 2005.

Seed Preparation Seeds of R. communis were obtained from Pharmacology Department of the University of Jos. These were deshelled and then crushed into fine powder using laboratory mortar and pestle.

Soxhlet Extraction The solvent used was absolute methanol. Twenty grams of the ground dry sample of the seed was placed in an extracting thimble and placed in the soxhlet apparatus. A water condenser was attached to the soxhlet apparatus at the top. The apparatus was fitted into the neck of a flask containing 250mls of the methanol (solvent) heated on a water bath.

The vapour from the solvent reached the soxhlet apparatus through the side tube and condensed on passing into the condenser. The condensed solvent dropped on the crude substance in the thimble and dissolved the required substance. The solution filtered through the thimble into the flask bearing the solvent. This process continued until the solvent from the thimble was colourless. Extraction was then said to be completed. This continuous extraction method extracted all the components of the plant which were soluble in methanol. The extract was then evaporated to dryness and a light brown oily extract was collected, weighed and stored by refrigeration at temperature of 40C for further susceptibility testing. Similar procedure was carried out for water extraction where water was used in place of methanol.

Antimicrobial Susceptibility Testing Bacteria used for the study were obtained from the Microbiology laboratory of the Jos University Teaching Hospital (JUTH), Jos. Organisms tested were: Klebsiella pneumoniae, Escherichia coli, Proteus mirabilis, Staphylococcus aureus, Pseudomonas aeruginosa and Enterococcus faecalis. Whatmann no 1 filter paper was used to prepare susceptibility discs of 4mm in diameter which were sterilized in hot air oven. Commercially prepared discs of ceftriaxone (30ug) were used as positive control which was susceptible against all the organisms tested, while either sterile distilled water or methanol was used as negative control. The refined oil extracts obtained was either mixed with methanol or warm sterile distilled water in varying concentrations. With a fine pipette, 0.02 mls of each concentration was impregnated with a sterile sensitivity disc; similar preparations were done with methanol and sterile distilled water. Sensitivity testing agar media were dried for 30 minutes at 370C and then flooded with about 0.5 McFarland's broth culture of the selected organisms. Using sterile forceps, the commercially prepared ceftriaxone discs and discs impregnated with appropriate concentrations of methanol and water extracts along with the negative controls were carefully placed on the flooded agar media.

The preparation was incubated overnight at 370C and the diameters (in millimeters) of zones of inhibition were measured using vernier calipers<sub>28</sub>.

Interpretation of Results The sensitivity report was interpreted as Sensitive (S), Intermediate (I) and Resistant (R) as follows:

Sensitive (S) Zone radius of inhibition wider than, equal to, or not more than 3mm smaller than the positive control.

Intermediate (I) Zone radius of inhibition is more than 3mm smaller than the positive control but not less than 3mm.

Resistant (R) No zone of inhibition or zone radius measures 2mm or less.

# RESULTS

All the organisms tested were Resistant to both the methanol and water extracts at 5mg/ml concentrations except Proteus mirabilis which was Intermediate against the water extract. Similarly, most organisms were Resistant to the extracts at 6mg/ml strengths except the water extracts against Proteus mirabilis (Intermediate), and methanol extracts against Staphylococcus aureus (Intermediate).

For the 7mg/ml concentrations, Proteus mirabilis and Staphylococcus aureus were Intermediate for both forms of extracts; Klebsiella pneumoniae Intermediate and Resistant for the methanol and water extracts respectively; Pseudomonas aeruginosa Resistant and Intermediate for the methanol and water extracts respectively. Both extracts were Resistant against Enterococcus faecalis but Intermediate and Sensitive against Escherichia coli respectively.

Most of the organisms were Intermediate to Sensitive for the 8mg/ml extracts of both methanol and water except Enterococcus faecalis which was Resistant to both extracts as well as the methanol and water extracts of Pseudomonas aeruginosa and Klebsiella pneumoniae respectively.

Both the methanol and water extracts at 9mg/ml strengths of the extracts were Intermediate to Sensitive against all the organisms tested, while the 10mg/ml of both extracts were Sensitive against all except the water extracts against Pseudomonas aeruginosa which was Intermediate.

#### Figure 1

Table 1: Antimicrobial susceptibility patterns of bacteria to various concentrations of the seed extracts of

Concentration of extracts inside disks* used (volume =0.02mls)												
	5mg/ml		6mg/ml		7mg/ml		8mg/ml		9mg/ml		10mg/ml	
	A	В	A	В	A	В	A	В	A	В	A	В
Klebsiella pneumoniae	R	R	R	R	Ι	R	Ι	R	Ι	Ι	S	S
Proteus mirabilis	R	Ι	R	Ι	Ι	Ι	S	Ι	S	Ι	S	S
Staphylococcus aureus	R	R	Ι	R.	Ι	Ι	Ι	Ι	Ι	S	S	S
Pseudomonas aeruginosa	R	R	R	R	R	Ι	R	Ι	S	Ι	S	Ι
Enterococcus faecalis	R	R	R	R	R	R	R	R	Ι	Ι	S	S
Escherichia coli	R	R	R	Ι	Ι	S	S	S	S	S	S	S

Key: A= Methanol Extracts B= Water Extracts S= Sensitive I= Intermediate

R= Resistant \*Disk Diameter= 4mm

# DISCUSSION

Generally, all the bacteria tested (Klebsiella pneumoniae, Escherichia coli, Proteus mirabilis, Staphylococcus aureus, Pseudomonas aeruginosa and Enterococcus faecalis) had a steady increase in susceptibility patterns from almost all resistant at 5mg/ml concentration through intermediate/sensitive at 9mg/ml to virtually all sensitive at 10mg/ml. This finding points to the fact that, there is a possibility to exploit the antibacterial properties inherent in the seed extract of R. communis for large scale medicinal uses. The finding is beneficial as it also heralds probably the emergence of a new antibiotic with such a wide spectrum of activity as found in the study being added to the existing list of them.

The active antibacterial ingredients in the Castor seed extracts should be identified and processed in possibly commercial quantities in order to seek its relevance in the current war against antimicrobial resistance. This no doubt poses a serious challenge to the modern day practice of medicine<sub>29,30,31,32</sub>. The fact that treatment of infections caused by organisms such as Staphylococcus aureus, Pseudomonas aeruginosa and Enterococcus faecalis are increasingly becoming difficult<sub>31,32</sub> further strengthens the importance of these present findings and the need for a continuous search of antibiotics with comparative advantage. Pseudomonas aeruginosa appeared less susceptible to the extracts compared to the other organisms. The high profile resistance of this organism against several antimicrobials in current use has severally been documented<sub>33</sub>,<sup>34</sup>.

Further work should be carried out to identify the active ingredients with the antibacterial properties as well as the tolerable human dose range vis-à-vis the minimum inhibitory concentration (MIC).

In conclusion, seed extracts of R. communis of both ethanol and water preparations were found to be highly active against several bacteria tested. Hence, active ingredients of these seed extracts should be identified and consequently its medicinal benefits to humans exploited.

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# References

1. De Vendra C, Raghavan G V. Agricultural by-products in South-East Asia: availability, utilization and potential value. World Rev Anim Prod 1978; 14(4): 11-27 2. Solaki R M, Bhadu V B, Jadav K V. Productivity of ground nut castor intercropping system as influenced by row ratio sewing time and hybrids of Castor, Ricinus communis L. J Oilseeds Res 2006; 23(2): 225-229. 3. Ahn Y J, Vang L, McKeon T A, Chen G Q. High frequency plant regeneration through adventitious shoot formation in castor (Ricinus communis L). In vitro cellular and Developmental Biology Plant 2007; 43(1): 9-15 4. Aslani M R, Maleki M, Moluri M, Shanti K, Najjar-Nezhad V, Afshan T. Castor bean (Ricinus communis) toxicosis in a sheep flock. Toxicom 2007; 49(3): 400-406. 5. Iyothi M, Thatikunta R, Akula B. Identification of castor, Ricinus communis L genotypes for rainfed conditions. J Oilseeds Research 2006; 23(2): 377-378. 6. Challener K R, McCarron M M. Castor bean intoxication. Ann Emerg Med 1990; 1177-1183. 7. Kinamore P A, Jaeger R W, de Castro F. Abrus and Ricinus ingestion: management of three cases. Clinical Toxicol 1980; 17(3): 401-405. 8. Venugopal C, Reddy G K, Reddy D S. Seed yield and not returns of rainfall Castor Ricinus communis L as influenced by plant geometry and nitrogen levels. J Oilseeds Research 2006; 23(2): 353-355. 9. Huguet-Termes T. New world materia medica in Spanish renaissance medicine: from

scholarly reception to practical impact. Med Hist 2001; by fluorescein-conjugated lectins. Z Parasitenkd 1985; 71(4): 443-458. 45(3): 359-376. 10. 22. Villalta F, Kierszenbaum F. Enhanced multiplication of 10. Wilcox M L, Bodeker G. Traditional herbal medicines intracellular (amastigotes) for malaria. B M J 2004; stages of Trypanosoma cruzi in vitro. J Protozool 1984; 41(3): 487-489. 23. Ng Y K, Ling E A. Emperipolesis of lymphoid cells in 329(7475): 1156-1159. 11. Varscht J, Tomos D, Komor E. Sugar concentrations vagal efferent neurons along and across the Ricinus communis L hypocotyl measured by single cell sampling following an intraneural injection of ricin into the vagus analysis. Planta 2006; nerve in rats. Neurosci Lett 224(6): 1303-1314. 1999; 270(3): 153-156. 12. Lakshminarayana M, Sujatha M.Toxicity of Bacillus 24. Jombo G T A, Egah D Z, Ayeni J A. Antibiotic susceptibility patterns of bacterial thuringiensis var kurstaki strains and purified crystal proteins against spodoptera litura isolates from urine samples of acquired immunodeficiency (Fabr) on castor, Ricinus syndrome (AIDS) patients in Jos, Nigeria. Mary Slessor J Med 2006; 6(2): 40-49. 25. Jombo G T A, Ayilara A O, Bello K, Gadzama G B, communis L. J Oilseeds Research 2005; 22(2): 433-434. Mbaawuaga E M. Patterns of bacterial isolates from surgical sites infections at a Federal 13. BoeckNeto R J, Gabrielli M F R, Shibli J A, Marcantonio E, Lia R C C, Marcantonio Medical Centre in North E. Histomorphometric evaluation of human sinus floor Eastern Nigeria. Mary Slessor J Med 2006; 6(2): 59-66. augmentation healing 26. Jombo G T A, Egah D Z, Banwat E B, Ayeni J A. responses to placement of calcium phosphate or Ricinus Nosocomial and community communis polymer acquired urinary tract infections at a Teaching Hospital in associated with autogenous bone. Clin Implant Dentistry & North Central Nigeria: Related Research 2005; Findings from a study of 12,458 urine samples. Nig J Med 7(4): 181-188. 2006; 15(3): 230-236. 14. Korwar G R, Pratibha G, Ravi V, Kumar D P. 27. Centres For Disease Control and Prevention (CDC). Performance of Castor (Ricinus Staphylococcus aureus resistant communis) and greengram (Vigna radiate) in agroforestry to vancomycin in United States, 2002. MMWR Morb Mortal Wkly Rep 2002; 51(26): systems in semi-arid tropics. Indian J Agronomy 2006; 51(2): 112-115. 565-567. 15. Isichei C O, Das S C, Ögunkeye Ö O, Okwuasaba F K, 28. Scott A. C. Laboratory control of antimicrobial therapy. Uguru V E, Onoruvwe O, et In: Mackie & al. Preliminary clinical investigation of the contraceptive McCartney Practical medical microbiology (Edited by, Collee JG, Duguid JP, Fraser efficacy and chemical pathological effects of RICOM- 1013-J of Ricinus AG and Marmion B P) 13th Edn. Vol 2, United Kingdom, communis var minor on women Edinburgh: Churchill volunteers. Phytother Res 2000; 14(1): 40-42. Livingstone, 1989; 161-181. 16. Das S C, Isichei C O, Okwuasaba F K, Uguru V E, 29. Albertin M T, Benoit C, Berardi L, Berrounane Y, Boisivon A, Cahen P, et al. Onoruvwe O, Olayinka A O, et al. Clinical pathological and toxicological studies of the effects Surveillance of methicillin-resistant Staphylococcus aureus of RICOM-1013-J of (MRSA) and Ricinus communis var minor on women volunteers and Enterobacteriaceae producing extended-spectrum betarodents. Phytother Res 2000; lactanase (ESBLE) in 14(1): 15-19. Northern France: a five year multicentre incidence study. J 17. Hetland G, Mollnes T E, Garred P. The B chain but not Hosp Infect2002; 52(2): 107-113. the A chain of Ricinus 30. Cuevas O, Cercenado E, Vindel A, Guinea J, Sanchezcommunis activates human complement. Cancer Lett 1993; 75(1): 59-63. Condi M, Sanchez-Somolinos M, et al. Evolution of the antimicrobial resistance of 18. Fakhri Z I. Mean wheal diameter in skin tests for castor bean extracts in castor bean Staphylococcus spp. in Spain: allergic workers of eastern Sudan. J Soc Occup Med 1989; Five nationwide prevalence studies, 1986 to 2002. 39(4): 144-146. Antimicrob Agents Chemother 19. Ikeda J, Kawakami H, Asano T, Takata K, Hirano H, 2004; 48(11): 4240-4245. Hirakawa K. Distribution of 31. Ishikawa K, Miyakawa S, Tanaka T, Naide Y, Shiroki R, glycoconjugates in gerbil hippocampal neurons-Hoshinaga K. The trend and susceptibility to antibacterial agents of Enterococcus spp. histochemical study with lectins. No To Shinkei 1991; 43(6): 539-543. from urinary tract 20. Ilavarason R, Mallika M, Venkataraman S. Antiinfections. Nippon Hinyokika Gakkai Zasshi. 2004; 95(1): inflammatory and free radical 25-34. 32. Bouza E, Garcia-Garrote F, Cercenado E, Marrin M, scavenging activity of Ricinus communis root extract. J Ethnopharmacol 2006; Diaz M S. Pseudomonas 103(3): 478-480. aeruginosa: a survey of resistance in 136 hospitals. 21. Choroma A, Ski L, Beat D A, Nordin J H, Pan A A, Antimicrob Agents Chemother. Honigberg B M. Further studies 1999; 43(4): 981-982. on the surface saccharides in Trichomonas vaginalis strains 33. Chastre J, Trouillet J L. Problem pathogens

(Pseudomonas aeruginosa and

Acinetobacter). Semin Respir Infect. 2000; 15(4): 287-298.

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