

# Antibacterial activity of herbal plant extracts towards the fish pathogens

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## Abstract

Fifteen plant extracts were screened against *Vibrio harveyi*, *V. parahaemolyticus* and *A. hydrophila* using disc method. Acetone extract of *Lawsonia inermis* Linn. had high antibacterial activity against three tested fish pathogens. Survival rate of 80% was observed when the *P. monodon* nauplii were exposed to the low concentration of 0.025 mg/ml in *L. inermis* extract. Due to its less toxic effect to shrimp and potent antibacterial activity the acetone extract of *L. inermis* could form one of the best options for developing novel antimicrobial compounds for ecofriendly management of disease caused by *V. harveyi*, *Vibrio parahaemolyticus* and *hydrophila*.

## INTRODUCTION

Shrimp farmers are loosing millions of dollars annually due to the large scale mortality of shrimp due to diseases caused by antibiotic resistance pathogens. Antibiotics resistance of microorganisms is a global concern (Austin, 1993).

Antibiotic resistant *V. harveyi* caused mass mortality in *Penaeus monodon* larvae in a hatchery (Karunasager et al., 1994). The frequency of resistance reflects the pattern of antibiotic usage. Plant secondary metabolites show activity in the micro to submicromolar range to Gram-positive species (Gibbons, 2004). Plant extracts decrease the selective pressure for developing antibiotic resistance (Lewis & Ausubel, 2006). Quinine and berberine from plants remain highly effective to fight against human microbial infections. Today plant materials are present in, or have provided the models for 50% of western drugs (Robbers et al., 1996) 5. Hence, in the present study plants containing antibacterial compound against *Vibrio harveyi*, *V. parahaemolyticus* and *A. hydrophila* were screened and their toxicity to shrimp, *Penaeus monodon* and *Artemia* were studied.

## MATERIAL AND METHODS

*Vibrio harveyi*, *V. parahaemolyticus* and *A. hydrophila* were obtained from the Central Institute for Brackishwater Aquaculture (CIBA), Chennai and maintained on Zobell's marine agar slants (Himedia, Mumbai). The agar-disc diffusion method of Bauer et al. (1966) was followed for testing the sensitivity of the isolates to 7 standard antibiotics against *Vibrio harveyi* on Mueller Hinton agar supplemented

with 2% sodium chloride. The plates were inoculated with 24 h cultures from Zobell marine broth. The discs were then placed on the agar plates and incubated at 30 to 37°C for 20 h.

The leaves of freshly harvested plants were washed and chopped with a knife to pieces. About 10g of chopped leaves were immersed in 50ml of acetone and kept in a shaker overnight at 100 rpm speed. The extract was filtered using Whatman no:1 filter paper. The filtrate was concentrated by keeping the extract in the oven at 40°C. 100 mg of concentrated extract was dissolved in 1ml of acetone. Briefly paper discs of 5mm diameter were prepared using Whatman filter paper (No:1) and sterilized. 5 l of plant extracts was impregnated on the discs (500 g/disc) and was dried at room temperature. In the same way control disc was also prepared by using acetone. Plants that showed antibacterial activity against *Vibrio harveyi*, *V. parahaemolyticus* and *A. hydrophila* were selected. Disc at different concentration of plant extracts (5 l, 10 l, 20 l, 40 l and 80 l) were prepared and dried at room temperature. In the same way control disc was also prepared by using acetone. Antibacterial activity was measured using disc diffusion method. *P. monodon*, *Artemia* nauplii were collected from a hatchery located in the east coast of India. They were maintained in the glass tank with aeration. They were separated into 5 groups and each containing 100 nauplii per bowl. The crude extract was added in each bowl at different concentrations (0.025, 0.05, 0.1, 0.2 and 0.4mg/ml). The extracts were prepared in 2% ethanol. The survival rate of the nauplii was observed after

24h.

## RESULTS AND DISCUSSION

Out of fifteen plant extracts screened, *L. inermis* and *T. indica* had antibacterial compound against *V. harveyi*, *Vibrio parahaemolyticus* and *Aeromonas hydrophila*. *Adhatoda vasica*, *Azadirachta indica* and *Nelumbium speciosum* showed antibacterial activity against *Vibrio parahaemolyticus* and *Aeromonas hydrophila* as seen in Table 1. Plants have broad spectrum antibacterial activity against human pathogens (Silva et al., 1997 and Navarro and Delgado, 1999). *V. cholerae* was sensitive to ethanol extract from *Terminalia macroptera* (Silve et al., 1997). Samy and Raja (1996) reported that *V. parahaemolyticus* and *V. damsela* were resistant to 16 aqueous plant extracts.

**Figure 1**

Table 1: Antibacterial activity of plant extracts against

Sl. No.	Plants	Family	Antibacterial activity		
			Vh	Vp	Ah
1	<i>Anacardium occidentale</i> , Linn.	Anacardiaceae	-	-	-
2	<i>Adhatoda vasica</i> Nees	Acanthaceae	-	+	+
2	<i>Anona squamosa</i> , Linn.	Annonaceae	-	-	-
3	<i>Azadirachta indica</i> , A. Juss.	Meliaceae	-	+	+
4	<i>Cardiospermum halicacabum</i> , Linn.	Sapindaceae	-	-	-
5	<i>Carica papaya</i> , Linn.	Caricaceae	-	-	-
6	<i>Clitoria ternatea</i> , Linn.	Fabaceae	-	-	-
7	<i>Coriandrum sativum</i> , Linn.	Umbelliferaceae	-	-	-
8	<i>Crataeva religiosa</i> , Forst.	Capparidaceae	-	-	-
9	<i>Hibiscus rosasinensis</i> , Linn.	Malvaceae	-	-	-
10	<i>Indigofera tinctoria</i> , Linn.	Fabaceae	-	-	-
11	<i>Lawsonia inermis</i> , Linn.	Lythraceae	+	+	+
12	<i>Murraya koenigii</i> , Spreng.	Rutaceae	-	-	-
13	<i>Nelumbium speciosum</i> , willd.	Nymphaeaceae	-	+	+
14	<i>Rosa damascena</i> , Mill.	Rosaceae	-	-	-
15	<i>Tamarindus indicus</i> , Linn.	Caesalpinaceae	+	+	+

**Figure 2**

Table 2: Antibacterial activity of plant acetone extracts against

Concentration (μl)	Zone of inhibition (mm)	
	<i>L. inermis</i>	<i>T. indica</i>
5	7.0 ± 0	6.0 ± 0
10	7.33 ± 0.47	6.67 ± 0.47
20	7.66 ± 0.47	7.33 ± 0.47
40	8.33 ± 0.47	10.33 ± 1.88
80	10.33 ± 0.47	12.33 ± 2.05

Each value is mean ± SD of three values

**Figure 3**

Table 3: Antibacterial activity of plant extracts against

Plant	Concentration (μl)					Rank
	5	10	20	40	80	
<i>A. indica</i>	8.0 ± 0	8.33 ± 0.47	9.33 ± 1.25	10 ± 0.82	10.33 ± 1.25	III
<i>A. vasica</i>	-	6.0 ± 0	6.33 ± 0.47	6.67 ± 0.47	7.33 ± 0.47	V
<i>L. inermis</i>	8.66 ± 0.47	9.67 ± 0.47	10 ± 0.82	10.33 ± 0.94	11.0 ± 0.82	I
<i>N. speciosum</i>	6.0 ± 0	6.66 ± 0.47	7.0 ± 0	7.33 ± 0.47	7.67 ± 0.47	IV
<i>T. indicus</i>	0.66 ± 0.47	7.33 ± 0.47	7.67 ± 0.47	8.33 ± 0.47	10.67 ± 0.47	II

Each value is mean ± SD of three values

**Figure 4**

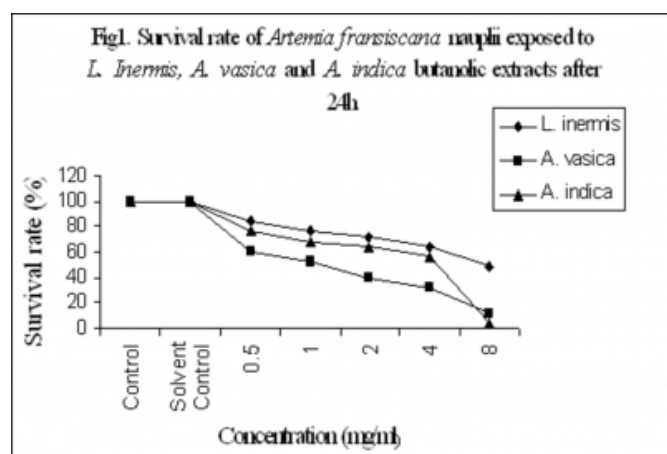
Table 4: Antibacterial activity of herbal plant extracts against

Plants	Control	Concentration (μl)					Rank
		5	1	2	4	8	
<i>A. vasica</i>	-	10 ± 1.0	11.66 ± 0.6	12.0 ± 0.0	13.33 ± 1.2	13.67 ± 0.6	II
<i>A. indica</i>	-	8.33 ± 0.6	8.66 ± 0.6	10.0 ± 0.0	11.0 ± 1.0	12.66 ± 0.6	III
<i>L. inermis</i>	-	10.67 ± 2.1	12.0 ± 2.2	13.67 ± 2.1	14.0 ± 1.0	15.33 ± 1.2	I
<i>N. speciosum</i>	-	-	6.33 ± 0.6	7.3 ± 0.6	8.0 ± 0.0	8.67 ± 0.6	V
<i>T. indicus</i>	-	7.67 ± 2.3	9.67 ± 2.3	10.33 ± 1.2	11.0 ± 1.4	11.33 ± 1.2	IV

Each value is the mean standard deviation of minimum three observations.

*L. inermis* showed high antibacterial activity against *V. harveyi* (Table 2). *V. parahaemolyticus* was sensitive to *Lawsonia inermis* with 11 mm inhibition zone as in Table 3. *L. inermis* extracts had high antibacterial activity against *A. hydrophilla* with antibacterial activity of 15.3mm at 80 l as in Table 4. Earlier reports showed the presence of bioactive compounds in *L. inermis*. The ethyl acetate extract of *L. inermis* showed broad spectrum of antibacterial activity (Ali et al., 2001). Tuberculostatic activity of the *L. inermis* was reported by Sharma (1990). Survival rate of 80% was observed at the concentrations of 0.025 mg/ml. of *L. inermis* extract. *L. inermis* extract was less toxic to *P. monodon* nauplii. The maximum survival rate of 48% was observed when the *Artemia* nauplii were exposed to *L. inermis* extract at 16 mg/ml after 24h(Fig.1).

Figure 5



Spray-dried preparation of *Tetraselmis suecica* inhibited shrimp pathogenic strains of *Vibrio* sp., *V. alginolyticus*, *V. anguillarum*, *V. parahaemolyticus* and *V. vulnificus* (Austin and Day, 1990). Veterinary pathogens have been inhibited by herbal medicines (Ernst, 1998). *Aloe vera* and *A. spicata* were used for chicken health management (Marizvikuru et al., 2005). *V. anguillarum* was inhibited by the dichloromethane extracts of *Asparagopsis armata*, *Ceramium rubrum*, *Drachiella minuta*, *Falkenbergia rufolanosa*, *Gracilaria cornea* and *Halopitys incurvus* (Bansemir et al., 2006). The pathogens *A. hydrophila* and *V. alginolyticus* were inhibited by *H. scoparia*, *L. acaciae*, and *P. harmala* (7-20.5 mm) (Bansemir et al., 2006). *A. hydrophila*, *P. fluorescens* and *Myxobacteria* spp. exhibited maximum sensitivity to Aquaneem in terms of percentage reduction of bacterial cell population in comparison to *E. coli* (Das et al., 1999). Due to its less toxic effect to shrimp and potent antibacterial activity the acetone extract of *L. inermis* could form one of the best options for developing novel antimicrobial compounds for ecofriendly management of disease caused by *V. harveyi*, *Vibrio parahaemolyticus* and *Aeromonas hydrophila*.

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