

Antifungal Activity of *Pseudomonas fluorescens* Against Different Plant Pathogenic Fungi

R Srivastava, Shalini

Citation

R Srivastava, Shalini. *Antifungal Activity of Pseudomonas fluorescens Against Different Plant Pathogenic Fungi*. The Internet Journal of Microbiology. 2008 Volume 7 Number 2.

Abstract

Antifungal activity of different strains of *Pseudomonas fluorescens* were tested against some plant pathogens such as *Alternaria cajani*, *Curvularia lunata*, *Fusarium* sp., *Bipolaris* sp. and *Helminthosporium* sp. in in vitro. Different concentrations (1000, 2000, 3000, 4000 and 5000 µg/mL) of *Pseudomonas fluorescens* were used and maximum spore germination of fungus was inhibited at 4000 and 5000 µg/mL. The result indicated that all the strains of *Pseudomonas fluorescens* presented a most significant value against *Alternaria cajani* and *Curvularia lunata*. Out of the five strains studied, the best result was shown by A-5, which showed almost complete inhibition against pathogenic fungi such as *Curvularia lunata* and *Fusarium* sp. at 4000 and 5000 µg/mL while strain L-5 was resistant against *Fusarium* sp. and *Helminthosporium* sp. at 5000 µg/mL. Among the fungus tested, bacterial strains C-03 and Pf4-1 were found to be more sensitive to *Fusarium* sp. and *Helminthosporium* sp.

INTRODUCTION

The surfaces of aerial plant parts provide a habitat for epiphytic micro-organisms, many of which also influence the growth of pathogens. Bacteria are generally the predominant initial inhabitants of newly expanded leaves, while yeasts and filamentous fungi dominate later in the growing season¹. A large body of information has been accumulated regarding antagonism between bacteria and fungi on the leaf surface, and its possible role in the biological control of pathogenic fungi². Biological control may be an alternative to chemicals in the control of some pathogenic fungi, or in order to reduce environmental pollution. Saprophytic organisms play an important part in reducing the incidence of foliar diseases from fungi and bacteria on crops in the field^{3,4}.

The genus *Pseudomonas* has been heterogenous since Migula first named it in 1984⁵. He designated and described the species associated with the genus in 1985⁶.

Pseudomonas are gram-negative, strictly aerobic, polarly flagellated rods. They are aggressive colonizers of the rhizosphere of various crop plants, and have a broad spectrum antagonistic activity plant pathogens, such as antibiosis (the production of inhibitory compounds)^{7,8}, siderophores production (iron-sequestering compounds) and nutrition or site competition⁹. Some species of *Pseudomonas*

can also produce levels of HCN that are toxic to certain pathogenic fungi¹⁰. These characteristics make *Pseudomonas* species good candidates for used as seed inoculant and root dips for biological control of soil-borne plant pathogen.

Pseudomonas fluorescens has the ability to grow at 4°C and hydrolyse gelatin. These characteristics help explain its frequent environment in spoilage of refrigerated food. The main property that conspires against its becoming important opportunistic pathogen is the inability to grow at body temperature. It is rarely pathogenic for humans, even though they have been found associated with empyema, urinary tract infections and septicemia. Some *Pseudomonas* have been recognized as antagonists of plant fungal pathogens and antibiotic producers¹¹. This is probably due to the abundance of this diverse group of bacteria and their obvious importance in the soils. *Pseudomonas* plasmids confer resistance to many antibiotics and antibacterial agents. Emmerich and Low¹² reported that the cell free culture of *Pseudomonas aeruginosa*, a concentrated to one tenth of its original volume, killed several kinds of bacteria. Due to the lytic action of culture broth on suspensions of some kinds of bacteria, they ascribed the inhibition to an enzyme termed pyocyanase. It has been used extensively in the therapy of diphtheria, influenza and meningitis¹³.

Antagonistic activity was also observed for *Pseudomonas* spp. in the rhizosphere has been recognized as a major factor in the suppression of many phytopathogens. Several antibiotic-like substances have been identified, including bacteriocins and phenazine antibiotics¹⁴, but one of the most important mechanisms responsible for the suppression of plant pathogens for *Pseudomonas* spp. is siderophore-mediated competitions for iron¹⁵. An inhibitory effect against pathogenic and spoilage bacteria by *Pseudomonas* species isolated from fish was also demonstrated¹⁶. Bacteria of the genus *Pseudomonas* comprise a large group of the active biocontrol strains as a result of their general ability to produce a diverse array of potent antifungal metabolites. These include simple metabolites such as 2,4-diacetylphloroglucinol, phenazine-1-carboxylic acid and pyrrolnitrin [3-chloro-4-(2'-nitro-3'-chlorophenyl)-pyrrole], as well as the complex macrocyclic lactone, 2,3-deoxy-2,3-didehydro-rhizoxin. Pyrrolnitrin is active against *Rhizoctonia* spp, *Fusarium* spp, and other plant pathogenic fungi, and it has been used as a lead structure in the development of a new phenylpyrrole agricultural fungicide¹⁷.

Strains of *Pseudomonas fluorescens* showed known biological control activity against certain soil-borne phytopathogenic fungi and has the potential to produce known secondary metabolites such as siderophore, HCN and protease that showed antagonistic activity against *Macrophomina phaseolina*, *Rhizoctonia solani*, *Phytophthora nicotianae* var. *parasitica*, *Pythium* sp. and *Fusarium* sp. Siderophore production in *Pseudomonas* strains inhibited the growth of *Staphylococcus*, *Escherichia coli* and *Aeromonas hydrophila* to 96.7%¹⁸. *Pseudomonas* spp. isolates from *Tuber borchii* ascocarps, known to be able to produce phyto-regulatory and biocontrol substances in pure culture, were used to perform studies on their possible physiological role in nature. Antimycotic activity was confirmed against fungal contaminants isolated from the ascocarps, suggesting that populations associated with *Tuber borchii* fruit bodies may play a role in the maintenance of ascocarp health. On the contrary, growth of the arbuscular mycorrhizal fungus *Glomus mosseae* and the ectomycorrhizal fungus *Laccaria bicolor*, putative competitors of *Tuber* for mycorrhizal infection sites on roots, was not influenced by the presence of any bacterial strain. The possibility that these bacteria, which show antifungal activity and fungal growth modulation activities, might be incorporated in the developing ascocarp by means of their preferential adhesion to *Tuber* mycelium¹⁹.

The purpose of this study was to examine the antifungal activity of *Pseudomonas fluorescens* against different plant pathogenic fungi viz. *Alternaria cajani*, *Curvularia lunata*, *Fusarium* sp., *Bipolaris* sp. and *Helminthosporium* sp. in vitro.

MATERIALS AND METHODS

ISOLATION OF CULTURES

Five strains of *Pseudomonas* which showed strong in vitro antifungal activity against some plant pathogens such as *Alternaria cajani*, *Curvularia lunata*, *Fusarium* sp., *Bipolaris* sp. and *Helminthosporium* sp. was isolated from different disease suppressive soils of King's B medium. The bacterial strains were cultured in nutrient agar at 28±2°C. For long-term maintenance the strains were preserved in nutrient broth containing 15% v/v glycerol at -70°C.

TEST ORGANISMS AND CULTURE MEDIA

Test fungi were isolated on potato dextrose agar (PDA) (peeled potato 250 g, dextrose 20 g, agar 15 g, distilled water 1 L) medium from their respective hosts collected from experimental farm of Banaras Hindu University, Varanasi, India. The cultures were further purified by single spore isolation technique and maintained at 25±2°C on PDA slants. 7-10 days old culture were used in the experiment. The fungi included in the present study are *Alternaria cajani*, *Curvularia lunata*, *Fusarium* sp., *Bipolaris* sp. and *Helminthosporium* sp.

Stock solution (5000 µg/mL) of the *Pseudomonas* strains was prepared by dissolving 5 mg of the culture in 1 mL of distilled water. Required concentrations (1000, 2000, 3000, 4000 and 5000 µg/mL) were prepared from each stock solution by diluting with distilled water. One drop (40 µL) from each concentration was placed on grease-free glass slides. Fungal spores (200-300) were picked up from 7-10 days old culture with sterilized inoculation needle and mixed in solution of the fraction of different concentrations separately. The slides were placed in moist chambers made by placing two sterile filter papers each on the lid and base of the petriplates. The slides with spores were then incubated at 25±2°C for 24 hr. Germination was observed after staining with cotton blue prepared in lactophenol under binocular microscope (Nikon, Japan Type 102). Spores mixed in sterile distilled water only served as control. All the experiments were conducted in triplicate.

RESULTS AND DISCUSSION

Five strains of *Pseudomonas fluorescens* were screened for

Antifungal Activity of *Pseudomonas fluorescens* Against Different Plant Pathogenic Fungi

their antimicrobial activity against different fungi viz. *Alternaria cajani*, *Curvularia lunata*, *Fusarium sp.*, *Bipolaris sp.* and *Helminthosporium sp.* in vitro. The *Pseudomonas* strains showed antifungal activity against all tested strains (Fig. 1 and 2). Table 1 presents the different concentrations obtained for each strain tested.

Figure 1

Table 1. Effect of on spore germination of some fungi

S.No.	Fungus	Host	Strains	Concentration ($\mu\text{g/ml}$)					
				Control	1000	2000	3000	4000	5000
1	<i>Alternaria cajani</i>	<i>Cajanus cajan</i>	A-5	33.33	50.00	78.57	86.21	87.50	92.86
			C-03	31.25	75.53	80.72	85.00	90.48	94.00
			CRM-3	23.81	57.78	82.14	85.25	86.67	94.29
			L-5	28.57	35.29	73.68	76.81	79.31	81.82
			PF4-1	33.93	64.76	65.33	73.91	87.84	90.28
2	<i>Curvularia lunata</i>	<i>Sorghum vulgare</i>	A-5	47.31	53.13	76.92	91.67	100.00	100.00
			C-03	27.27	73.68	80.88	91.21	91.46	94.64
			CRM-3	31.75	67.65	70.00	83.78	84.21	91.11
			L-5	37.14	48.28	53.33	63.16	75.00	84.62
			PF4-1	40.91	75.00	75.00	83.64	87.67	89.71
3	<i>Fusarium sp.</i>	<i>Cajanus cajan</i>	A-5	23.30	75.18	88.89	95.08	97.75	100.00
			C-03	8.89	17.07	18.37	21.57	22.54	26.67
			CRM-3	23.26	65.38	66.67	70.00	81.82	90.70
			L-5	50.61	66.67	86.89	88.14	89.47	97.94
			PF4-1	14.89	15.79	17.34	18.18	20.69	22.73
4	<i>Bipolaris sp.</i>	<i>Oryza sativa</i>	A-5	49.57	71.79	85.19	87.50	95.92	96.43
			C-03	18.73	35.48	55.17	58.33	62.82	73.27
			CRM-3	10.64	25.71	30.77	35.14	54.29	59.26
			L-5	6.59	10.53	12.07	23.53	26.67	27.27
			PF4-1	34.21	61.11	81.25	86.84	94.59	95.65
5	<i>Helminthosporium sp.</i>	<i>Saccharum officinarum</i>	A-5	10.27	35.71	69.64	71.43	72.22	73.68
			C-03	17.86	20.00	23.40	23.68	28.30	33.33
			CRM-3	12.12	25.37	28.89	39.39	41.38	50.00
			L-5	77.78	79.41	88.14	91.43	94.92	100.00
			PF4-1	17.02	25.00	27.12	27.12	28.07	31.58

Figure 2

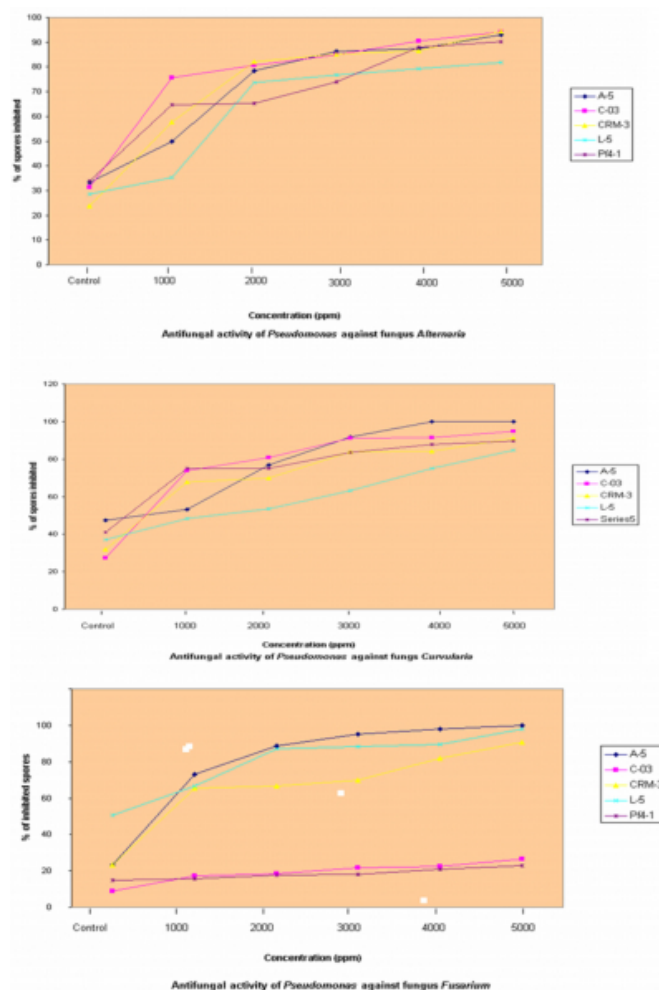
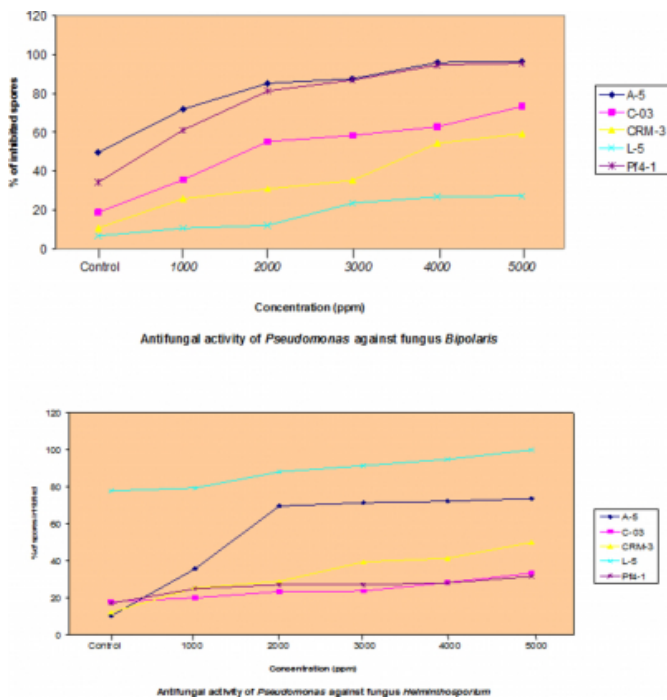


Figure 3



All the five strains showed varied levels of antifungal activity and the highest concentration of *Pseudomonas* strains were capable of inhibiting the growth of the pathogenic microorganism. At 5000 µg/mL, all the *Pseudomonas* strains (A-5, C-03, CRM-3, L-5 and Pf4-1) were highly resistant and showed highest inhibition percentage (81% to 100%) against fungus *Alternaria cajani* and *Curvularia lunata*. Out of the five strains studied, the best result was shown by A-5, which showed almost complete inhibition and maximum activity against *Curvularia lunata* and *Fusarium* sp. at 4000 and 5000 µg/mL while strain L-5 was resistant against *Fusarium* sp. and *Helminthosporium* sp. at 5000 µg/mL. The moderate activity was shown by the strains C-03 and CRM-3 against *Bipolaris* and A-5 and CRM-3 against *Helminthosporium*. The most sensitive strains were C-03 and Pf4-1 against pathogenic fungi *Fusarium* sp. and *Helminthosporium* sp. while strain L-5 against *Bipolaris* sp. at 5000 µg/mL. These data revealed that *Pseudomonas* strains exhibited significant antifungal activity. In testing, percentage of spores inhibition increased with increase in concentration and thus exhibiting concentration dependent activity.

The biological control of soil-borne pathogens with antagonistic bacteria, particularly *Pseudomonas* spp. belonging to plant growth promoting Rhizobacteria, has received prominent attention because of the dual role of these bacteria in plant-growth promotion and disease control

²⁰. When identifying potential biocontrol agents, antifungal metabolites produced by them or ability of these agents to induce antifungal compounds in plants are important factors to be taken into account. Many research groups are actively trying to find metabolites produced by biocontrol agents or induced by them in plants, and that will suppress particular diseases ²¹. In earlier studies ^{22,23} a talc-based formulation of the *P. fluorescens* strains also used here reduced sheath-blight incidence under field conditions in different zones of Tamil Nadu state, India.

Pseudomonas strains were evaluated for their ability to control *Sclerotinia homeocarpa* and *Bipolaris sorokiniana* on the phylloplane of Kentucky bluegrass ²⁴. *Pseudomonas*, *Bacillus* and *Stenotrophomonas maltophilia*, showed antifungal activity against *Verticillium dahliae* var. *longisporum* in vitro and were evaluated as potential biocontrol agents by Berg et al. ²⁵. Rajappan and Ramaraj ²⁶ evaluated the efficacy in vitro of *P. fluorescens* against the cauliflower wilt pathogen *Fusarium moniliforme*. Fluorescent pseudomonad strains found to be effective against *Sclerotium rolfsii* were evaluated by Patil et al. ²⁷ under greenhouse conditions for their effects on groundnut and on collar rot incidence. *Trichoderma viride* and *Pseudomonas* sp. controlled stalk rot (associated with *Pythium aphanidermatum* and *Fusarium graminearum*) at the seedling stage of maize ²⁸. Experiments in vitro and in vivo by Rozsnyay et al. ²⁹ showed that some strains of *P. fluorescens*, some epiphytic bacteria and some fungi inhibited canker and dieback diseases of apricot. Janisiewicz and Roitman ³⁰ reported that blue mold and grey mold of apples and pears could be controlled by *Pseudomonas*. A bacterial strain identified as *Pseudomonas acidovorans* NB-10II which has been renamed *Comamonas acidovorans* ³¹, isolated from water pond in South Jordan was found to have an antifungal activity against filamentous fungi (*Aspergillus niger* SQ 40, *Fusarium oxysporium* SQ 11, *Verticillium dahliae* SQ 42), yeasts (*Saccharomyces cerevisiae* SQ 46, *Candida albicans* SQ 47). This bacterial isolate was found to accumulate the main portion of the antimicrobial substances in their cells ³².

Recent increases in fungal infections, the few available antifungal drugs, and increasing fungal resistance to the available antifungal drugs have resulted in a broadening of the search for new antifungal agents. Strains of *Pseudomonas syringae* pv. *Syringae* produce cyclic lipopeptidoneptides that showed broad antifungal activity and fungicidal actions. Overall, the cyclic

lipodepsinonapeptides were more effective against yeasts than against the filamentous fungi³³. *Pseudomonads* represents the major group of non-differentiating microorganisms that produce antibiotics such as phycocyanin, pyrrolnitrin and pseudomonic acid, was investigated in in vitro and in vivo that showed anticandidal activity against *Candida* species³⁴.

CONCLUSION

The presented data exhibit the antifungal activity of *Pseudomonas* strains and indicate the possibility of using *Pseudomonas fluorescens* as a biological control agent of some plant pathogenic fungi. However, this requires further screening of a large number of *Pseudomonas* strains from different regions of India. The antimicrobial activity of *Pseudomonas* may be attributed to the various phytochemical constituents have even more potency with respect to the inhibition of microbes.

ACKNOWLEDGEMENTS

We are grateful to the Institute of Bioengineering and Biological Sciences, Varanasi, India to provide financial support and lab facilities to carry out the present investigation.

References

1. Kinkel, L.L., J.H. Andrews, F.M. Berbee and E.V. Nordheim. Leaves as islands for microbes, *Oecologia* 71: 405–408. 1987
2. Gowdu, B.J. and R. Balasubramanian. Role of phylloplane micro-organisms in the biological control of foliar plant diseases, *Zeitschrift Pflanzkrankheit und Pflanzenschutz* 95(3): 310–331. 1988.
3. Blakeman, J.P. and N.J. Fokkema. Potential for biological control of Plant diseases on the phylloplane, *Annual Review of Phytopathology* 20: 162–192. 1982.
4. Frommell, M.I. and G. Pazos. Tomato rhizosphere and phyllosphere bacteria as potential biocontrol agents for fungal pathogens, *Phytopathologia Mediterranea* 28: 45–54. 1993.
5. Migula, W. *Arbeiten aus dem Bakteriologischen Institute der Technischen Hochschule Zu Karlsruhe* 1: 235-238. 1984.
6. Migula, W. *Bacteriaceae (Stabchenbakterien). Die Natürlichen Pflanzenfamilien.* Ehglar A, Prantl N (eds). Teil I, Abt La, W. Engelmann publishers. Leipzig. Pp 20-30. 1985.
7. Cartwright, D.K., W.S. Chilton, D.M. Benson. Pyrrolnitrin and phenazine production by *Pseudomonas cepacia*, strain 5.5 B, a biological agent of *Rhizoctonia solani*, *Appl. Microbiol. Biotechnol.* 43: 211-121. 1995.
8. Rosales A.M., L. Thomashow, R.J. Cook, T.W. Mew. Isolation and identification of antifungal metabolites produced by rice-associated antagonistic *Pseudomonas* spp., *Phytopathology* 85: 1028-1032. 1995.
9. Winkelmann, G. and H. Drechsel. Microbial siderophores. *Biotechnology.* HJ Rehm and G. Reed (eds.). Second Edition. VCH, Weinheim Vol. 7, Pp. 199-246. 1997.
10. David, N.D. and F. O’Gara. Metabolites of *Pseudomonas* involved in the biocontrol in the biocontrol of plant diseases, *Tibtech* April. 12: 133-141. 1994.
11. O’Sullivan D.J. and F. O’Gara Traits of fluorescent *Pseudomonas* spp. Involved in suppression of plant root pathogens, *Microbiol. Rev.* 56: 662-676. 1992.
12. Emmerich R. and D. Low. DBakteriolytische Enzyme als Ursache der erworbenen Immunität und die Heilung von Infektionskrankheiten durch dieselben, *Z. Hyg. Infektionskranken* 31: 1-65. 1899.
13. Leisinger T. and R. Margraff . Secondary metabolites of the fluorescent *Pseudomonads*, *Microbio. Rev.* 43: 422-442. 1979.
14. Hamdan H., D.M. Weller and L.S. Thomashow. Relative importance of fluorescent siderophores and other factors in biological control of *Gaeumannomyces graminis* var. *tritici* by *Pseudomonads fluorescens* 2-79 and M4-80R, *Applied and Environmental Microbiology* 57: 3270-3277. 1991.
15. Henry M.B., J.M. Lynch and T.R. Farmor. Role of siderophores in the biocontrol of *Pseudomonas tolaasii* by fluorescent pseudomonad antagonists, *Journal of Applied Bacteriology* 70: 104-108. 1991.
16. Gram L and J. Melchiorson. Interaction between fish spoilage bacteria *Pseudomonas* sp. and *Shewanella putrefaciens* in fish extracts and on fish culture, *Journal of Applied Bacteriology* 80: 589-595. 1996.
17. Ligon J.M., D.S. Hill, P.E. Hammer, N.R. Torkewitz, D. Hofmann, H.J. Kempf, K.H. van Pée . Natural products with antifungal activity from *Pseudomonas* biocontrol bacteria, *Pest Management Science* 56: 688 – 695. 2000.
18. Ahmadzadeh M, H. Afsharmanesh, M. Javan-Nikkhah, A. Sharifi-Tehrani. Identification of some molecular traits in fluorescent pseudomonads with antifungal activity, *Iranian Journal of Biotechnology* 4: 245-253. 2006.
19. C. Sbrana, G. Bagnoli, S. Bedini, C. Filippi, M. Giovannetti, and M. P. Nuti. Adhesion to hyphal matrix and antifungal activity of *Pseudomonas* strains isolated from *Tuber borchii* ascocarps, *Can. J. Microbiol.* 46(3): 259–268. 2000.
20. Zehnder G., J.F. Murphy, E.J. Sikora and Kloepper. Application of rhizobacteria for induced resistance, *European Journal of Plant Pathology* 107: 39-50. 2001.
21. Dowling D.N. and F. O’Gara. Metabolites of *Pseudomonas* involved in the biocontrol of plant disease, *Trends in Biotechnology* 12: 133-141. 1994.
22. Nandakumar R., S. Babu, R. Viswanathan, T. Raguchander and R. Samiyappan. Induction of systemic resistance in rice against sheath blight disease by *Pseudomonas fluorescens*, *Soil Biology and Biochemistry* 33: 603–612. 2001a
23. Nandakumar R., S. Babu, R. Viswanathan, J. Sheela, T. Raguchander and R. Samiyappan. A new bio-formulation containing plant growth promoting rhizobacterial mixture for the management of sheath blight disease and enhanced grain yield in rice, *Biocontrol* 46: 493–510. 2001b.
24. Hodges C.F., D.A. Campbell and N. Christians. Potential biocontrol of *Sclerotinia homoeocarpa* and *Bipolaris sorokiniana* on the phylloplane of *Poa pratensis* with strains of *Pseudomonas* spp., *Plant Pathology* 43(3), 500–506. 1994.
25. Berg G., P. Marten and H. Bahl. Population dynamics of bacteria including antifungal species in the rhizosphere of

- oilseed rape during its life cycle, Archives of Phytopathology and Plant Protection 31(3), 215–224.1998.
26. Rajappan K. and B. Ramarej. Evaluation of fungal and bacterial antagonists against *Fusarium moniliforme* causing wilt of cauliflower, Annals of Plant Protection Sciences 7: 205-207. 1999.
27. Patil R., K. Jagadeesh, P. Krishnaraj and J. Kulkarni. Bacterization of groundnut with *Pseudomonas fluorescens* for the control of collar rot caused by *Sclerotium rolfsii* Sacc. Karnataka, Journal of Agricultural Sciences 11(2): 423–425.1998.
28. Chen J., H.M. Gao, R.M. Lin, M.S. Ji and Z.G. Gao. Infection mechanism and biocontrol of major corn fungal diseases in Northern China, Research Progress in Plant Protection and Plant Nutrition. 78–84.1999.
29. Rozsnyay Z.D., M. Hevesi, Z. Klement and L. Vajna. Biological control against canker and dieback diseases of apricots, Acta Phytopathologica Entomologica Hungarica 27: 551-556. 1992.
30. Janisiewicz W.J. and J. Roitman. Biological control of blue mold and gray mold on apple and pear with *Pseudomonas cepacia*, Phytopathology 78: 1697–1700.1988.
31. Tomaoka J, D. Ha, K. Komagata. Reclassification of *Pseudomonas acidovorans* den Dooren de Jong 1926 and *Pseudomonas testoteroni* Marcus Talalay 1956 as *Comamonas acidovorans* comb nov. and *Comamonas testoteroni* comb nov., with an emended description of the genus *Comamonas*, Int. J. Syst. Bacteriol. 37: 52-59. 1987.
32. Nasse M and E.L. Banna. Antifungal activity of *Comamonas acidovorans* isolated from water pond in south Jordan, African Journal of Biotechnology 6: 2216-2219. 2007.
33. Sorensen K.N., K.H. Kim and J.Y. Takemoto. In vitro antifungal and fungicidal activities and erythrocyte toxicities of cyclic lipodepsinonapeptides produced by *Pseudomonas syringae* pv. *Syringae*, Antimicrobial Agents and Chemotherapy 40: 2710-2713. 1996.
34. Kaleli I, Cevahir N, Demir M, Yildirim U and Sahin R. Anticandidal activity of *Pseudomonas aeruginosa* strains isolated from clinical specimens. Mycoses 50: 74-78. 2007.

Author Information

Rachana Srivastava, Ph.D

Scientist, Department of Medicinal Chemistry, Institute of Bioengineering and Biological Sciences

Shalini, Ph.D

Scientist, Department of Biotechnology, Institute of Bioengineering and Biological Sciences