Antimicrobial Potentials of Yaji-Spices: The Constituents of a Complex Nigerian Suya Meat Sauce Inducing Histological Investigations

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Citation

A Nwaopara, C Anibeze, F Akpuaka, S Nwaopara. *Antimicrobial Potentials of Yaji-Spices: The Constituents of a Complex Nigerian Suya Meat Sauce Inducing Histological Investigations*. The Internet Journal of Alternative Medicine. 2008 Volume 6 Number 2.

Abstract

Spices and herbs have been used for thousands of centuries as preservatives for foods and for medicinal purposes. Some of these spices and herbs possess antimicrobial potentials that may, in combination, be considered as alternatives to conventional antimicrobial agents especially in this era of antimicrobial drug resistance. Our interest however, is on the widely consumed Nigerian suya meat sauce called Yaji. It is a complex mixture of salt, ginger, clove, red pepper and black pepper, all of which, on individual basis, have established antimicrobial properties. This paper therefore highlights these all-in-one antimicrobial potentials of Yaji.

INTRODUCTION

Despite the presence of biologically active substances, which, at high levels of consumption, can exhibit toxicity [1], spices have been included amongst the substances classified as nutrients and nutraceuticals [2]. In fact, it has been stated that tradition attaches all manner of benefit to every spice, condiment and herb as exemplified in the pharmacopoeias of Indian system of medicine including Ayurvedic, Sidha and Unani systems [324,576].

Worthy of note also is the fact that the 'science of nutrition' has been identified as a significant part of preventive medicine $[\,_{7}]$ and dietary regulations is an important component of the treatment of diabetes mellitus $[\,_{8}, 9, 10, 11]$, atherosclerosis $[\,_{12}, 13]$, constipation $[\,_{14}, 15, 16, 17]$ and other ailment associated with overweight $[\,_{18}, 19]$ and cancer $[\,_{20}, 21, 22]$. Indeed, researchers believe that with diligent efforts, herbs (and spices) can be utilized for the utmost advantage to humans $[\,_{23}, 24]$ and one such advantage, is their use as antimicrobial agents.

Antimicrobial agents are those substances or drugs with antifungal, antiviral, antiparasitic and antibiotic properties. According to the World Health Organization [25] antimicrobial agents have substantially reduced the threat posed by infectious diseases since their discovery in the 20th century. The use of these "wonder drugs", combined with

improvements in sanitation, housing, and nutrition, and the advent of widespread immunization programmes, has led to a dramatic drop in deaths from diseases that were previously widespread, untreatable, and frequently fatal. Over the years, such agents have saved the lives and eased the suffering of millions of people and have helped in bringing many serious infectious diseases under control. These drugs have also contributed to the major gains in life expectancy experienced during the latter part of the last century.

However, these gains are now seriously jeopardized by another recent development: the emergence and spread of microbes that are resistant to cheap and effective first-choice, or "first-line" drugs. As a result of this, it has been advised that the usage of antimicrobial agents by individuals is preferable when prescribed by a qualified physician since an overuse (or under-use), can propagate resistant microbial variants $[_{26,27,28}]$ and antimicrobial resistance is resulting in increased morbidity, mortality, and health-care costs $[_{29}]$.

Interestingly, scientific experiments since the late 19th century, have documented the antimicrobial properties of some spices, herbs and their components [30,31,32,33,34]. The interest in the use of compounds derived from spices as antimicrobial agents was sparked up by the changes in consumer attitudes towards the use of preservative agents such as nitrates and sodium chloride (NaCl) in foods during

in 1980s [31].

In this paper therefore, we highlight the antimicrobial potentials of the spices-in-combination within Yaji (a complex Nigerian meat sauce), since antimicrobial drug alternatives are becoming inevitable especially now that antimicrobial drug-dosage-compliant problems as well as 'self medication' is inducing microbial resistance against conventional antimicrobial agents.

Yaji is a complex mixture of groundnut powder, additives, spices and salt [35,36]. The spices in Yaji are ginger, cloves, red pepper, and black pepper, while the common additive other than salt, is Maggi [37]. Historically, the name 'Yaji' was an adaptation of the name of a 14th century Hausa ruler called "Yaji (meaning the 'hot one')" [38].

Yaji is a sauce for the meat delicacy called Suya; a boneless lean meat of mutton; beef or goat meat staked on sticks, coated with its sauce, oiled and then roasted around a burning fire [36]. According to Igene and Mohammed [39], Suya as a popular, traditionally processed, ready to eat Nigerian meat product, that may be served or sold along streets, in club houses, at picnics, parties, restaurants and within institutions. Omojola et al. [40] adds that Suya is one of such intermediate moisture product that is easy to prepare and highly relished while Uzeh et al. [41] described Suya as a mass consumer fast food whose preparation and sales along streets are usually not done under strict hygienic condition because they are still done locally.

Considering its mass-consumption rate and complexity, Yaji has become the basis for several histological investigations with interesting findings [37,42,43,44]. These investigations were aimed at determining the effect of excessive consumption of Yaji (or its constituent spices) on the Pancreas $[_{37}]$, Liver $[_{42}]$, Kidney $[_{43}]$ and the Heart $[_{44}]$. Available results show that an excessive consumption of Yaji-spices especially in combination, is capable of inducing Pancreatic, Liver and Kidney tissue damage [37,42,43] but with no significant effect on the histology of the Heart under the same experimental conditions [44]. However, one other aspect of the ongoing investigation on Yaji is the nonexperimental but analytical review of the potentials of Yaji that might be of public health concern; especially when one considers the large consumption population. Such is the report on the potential health hazards of Yaji by Nwaopara et al [45].

Judging from the histological findings above, one might conclude that the consumption of Yaji is always likely to produce a negative effect, which however, should not be the case, because the emphasis in the experiments was on the effect of an excessive consumption. In fact, we are convinced that a moderated consumption of Yaji has the potential to produce positive results in some sense and such can be seen in its antimicrobial potentials; considering the available information on the antimicrobial properties of its constituent spices.

THE ANTIMICROBIAL POTENTIALS OF

Some plant products can be considered to be natural sauces of antimicrobial agent as researchers from Ohio Weslyan University have reported that some birds select nesting material rich in antimicrobial agents and this was believed to protect their young from harmful bacteria [46]. There is also a report that lowland gorillas take 90% of their diet from the fruits of Aframomum melegueta, a relative of the ginger plant, which is a potent antimicrobial and apparently keeps Shigellosis and similar infections at bay [47]. Indeed, it has been shown that sick animals tend to forage plants rich in secondary metabolities, such as tennins and alkaloids [48] and these phytochemicals often have antiviral, antibacterial, antifungal and antihelminthic properties [47]. It has also been observed that higher levels of spices are necessary to inhibit microbial growth in food than in the culture media [31].

This reminds one about the letter dated 9th October 1676, in which Van Leeuwenhoek described the decline in the number and activity of "animalcules" in a sample of well water following the addition of pepper [49]. In 1996, Cichewicz and Thorpe [50] performed a survey of the Mayan herbal pharmacopoeia and found a number of herbal remedies directed at ailments probably of microbial origin. Most of the remedies list capsicum fruit, alone or in conjunction with the leaves, roots, or seeds as the main ingredient and these were applied to a variety of ailments including respiratory problems, bowel complaint, earaches and sores [50].

There is a report that ginger has strong antibacterial and to some extent antifungal properties [51]. In vitro studies have shown that active constituents of ginger inhibit multiplication of colon bacteria [51]. Ginger inhibits the growth of Escherichia coli, Proteus sp, Staphylococci, Strepto–cocci and Salmonella [52:53:54:55]. Hence, ginger should have impact on the growth of Bacillas cereus, which

mainly causes diarrhoea and nausea [54,55].

Ginger extract and its pungent compounds demonstrated greater antibacterial activity against a variety of bacteria species including Helicobacter pylori, Staphylococcus aureus, Pseudomonas aeruginosa and Escheroichia coli, although mixed result is attributed to different ginger preparations and varying strength [56]. Such an extract of ginger has antimicrobial action at levels equivalent to 2000 mg/ml of the spice [51]. In addition, ginger inhibits aspergillus, a fungus known for production of aflatoxin, a carcinogen [57,558]. Fresh ginger juice showed inhibitory action against A.niger, S.cerevisiae, Mycoderma SPP and L. acidophilus at 4, 10, 12 and 14% respectively at ambient temperatures [50].

The antimicrobial effect of clove is attributed to eugenol, which is the major active constituent of its essential oil $[a_0]$. The microbial inhibition of eugenol might be related to the membrane disruption or, according to Wendakoon and Sakaguchi [61], by inactivation of enzymes and genetic materials. The bactericidal activity of clove against food borne pathogens, like Listeria mono cytogenes was reported in TSB by Ting and Deibel [62] and in saline solution by Aureli et al. [63]. The results presented by Smith-Palmer et al. [64] established that the essential oil of clove retained their low bacteriostatic and bactericidal concentrations even at 40C. Stecchins et al. [65], suggested a marked reduction in the number of Aeromonas hydrophila in meat samples treated with clove oil. Clove extract at 2000Ngml-1 showed bactericidal activity towards Yersinia enterocolitica in TSB $[_{66}].$

It has been suggested that the cytoplasmic membrane is also target for eugenol action and results evidencing the kt efflux corroborated this hypothesis. This result was in agreement with Degre and Sylvestre [67] who considered that the probable antimicrobial activity of eugenol was on cellular lipids resulting in the loss of intracellular contents.

Capsicum baccatum, Capsicum chinese, Capsicum frutescens and Capsicum pubescens varieties were tested for their antimicrobial effects with fifteen bacterial species and one yeast specie [50]. Two pungent compounds found in Capsicum species (Capsaicin and Dihydrocapsaicin) were also tested for their antimicrobial effects [50]. The plain and heated extracts were found to exhibit varying degrees of inhibition against Bacillus cereus, Bacillus subtilis Clostridium sporogenes, Clostridium tetani, and

Streptococcus pyogenes [50].

Capsicum extracts were shown to be growth-inhibitory on three of the sixteen microbial species tested. Clostridium species are common anaerobic bacteria often found in soil, sewage, aquatic sediments and decaying organic matter, as well as in the intestines of animals $[_{68}]$. Clostridium tetani is the bacterium responsible for Tetanus and Clostridium botulinum (not tested) causes Botulism. Streptococcus pyogenes is the central pathogen identified in a variety of cutaneous and systemic infections. Thus, capsicum extracts do appear to have some valuable antimicrobial activity [68]. In fact, a later study has shown that Capsaicin strongly inhibits the growth of Bacillus subtilis [69] as aqueous and ethanolic extracts of black pepper have been screened for antibacterial activities against Penicillin G resistant strain of Staphylococcus aureus [70], Bacillus cereus and Bacillus subtilis [71].

Adebesin et al. [72] had observed in their study comparing the microbial counts of several peanut products, that there was a lower microbial count in Yaji, which they attributed to the antimicrobial potentials of its constituent spices. Of course available evidence supports the fact that spices inhibit microbial growth [30,33,34]. Again, the result of histological study conducted by Nwaopara et al. [43] on the Kidney showed the presence of round basophilic bodies in the interstitium of the kidney of rabbits following an excessive ingestion of Yaji spices. The argument was that in as much as the existing scientific evidence suggests that the presence of basophilic inclusion bodies in the kidney can be associated with microorganisms as reported in birds by Bernier et al. [73] and in humans by Nickeleit et al. [74,75] and Drachenberg et al. [76], there was an obvious probability that the distinct round basophilic bodies in the interstitium of the renal cortex of the kidney as observed in the experiment, might not be micro - organism - related considering the overwhelming evidence about the antimicrobial potential of the constituent spices in Yaji [52,53,57,58,59,77,78,79,80,81,82].

CONCLUSION

Based on the forgoing therefore, the possibility of Yaji becoming an antimicrobial agent is not in doubt considering the barrage of information that are available to substantiate the antimicrobial potentials of its spice-constituents. In fact, the issues of dosage compliance and drug resistance, as well as the 'faking of spices' as compared to the 'faking of conventional antimicrobials' are significantly ruled out.

Moreover, spices are readily available, cheap, and durable; and a regular intake can as well serve as a preventive measure. However, the moderation of Yaji consumption is preferable and safer.

ACKNOWLEDGEMENT

We express our appreciation to the authors of several publications from which we derived the information for this presentation. To these authors, we say thank you for your contribution to humanity.

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References

- Southgate DAT. Spices and Beverages in Essential Human Nutrition and Dietitics. Edinburgh, Churchill Livingstone, 1993. pp. 325 - 334.
 Hendrich S, Lee KW, Xu X, Wang HJ, Murphy PA.
- 2. Hendrich S, Lee KW, Xu X, Wang HJ, Murphy PA. Defining food components as new nutrition. Journal of Nutrition (1994); 124 (9 suppl): 1789s 1792s.
- 3. Charaka. Charak Samhita, Sutra Sthan. Varanasi, Chaukamba Surbharati Prakashan 1994; 18(32)353.
- 4. Pruthi JS. Spices and Condiments. New Delhi National Book Trust India, 1987; 5(41-43): 185-191.
- 5. Bhat NS, Bhat AD. Indegenous drugs and liver disease. Indian Journal of Gastroenterology 1996; 15(2): 63 67. 6. Hameed HA. The complete book of home remedies. New
- Delhi. Orient Paperbacks, 1982. pp 39 50. 7. Kochhar KP. Effects of Dietary Herbs and Spices. Journal of Authormolecular Medicine 1999; 14(4): 210 - 218.
- 8. Wolever TM, Nguyen P, Chaisson J, Junt JA, Josse RG, Palmason C, Rodger NW, Ross SA, Ryan EA, Tan MH. Determinants of diet Glycemic Index calculated retrospectively from diet records of 342 individuals with non-insulin dependent diabetes mellitus. American Journal of Clinical Nutrition 1994; 59:1265-1269.
- 9. Wolever TMS, Nutall FQ, Lee R. Prediction of the relative blood glucose response of mixed meals using white bread glycaemic index. Diabetes Care 1985; 8: 418 428. 10. Jenkins DJA, Wolever TMS, Jenkins AL. Modern Nutrition in Health and Disease. Philadelphia. Lea and Febiger 1994. pp. 583 602. 11. Anderson JW, Geil PB. Modern Nutrition in Health and
- 11. Anderson JW, Geil PB. Modern Nutrition in Health and Disease. Philadelphia. Lea and Febiger. (1994). pp. 1259 1286.
- 12. Abrams ME, Jareth RJ, Keen H., Boyns DR., Corsslay JN. Oral glucose tolerance and related factors in a normal population sample II, interrelationships of glycerides, cholesterol and other factors with glucose and insulin response. British Medical Journal 1969; 1: 599 602.

 13. Chait A, Brunzell JD, Denke, MA et al. Rationale of the Diet. Heart statement of the American heart Association Report of the Nutrition Committee. Ciculation 1993; 88(6):3008 3029.
- 14. Kurpad AV, Shetty PS. Methods of the estimation of intestinal transit time in the tropics. Tropical

- Gastroenterology 1985; 6:197 -202.
- 15. Welch MCL, Read NW. Regulation of gastrointestinal function by Ileal nutrients in health and disease. Tropical Gastroenterology 1986; 116 125.
- 16. Horowitz M., Wishart J, Medolox A, Russo A. The effect of Chilli on Gastrointestinal transit. Journal of Gastroenterol Hepatol 1992; 7(1): 526.
- Gastroenterol Hepatol 1992; 7(1): 526. 17. Holzer HH, Turkelson CM, Solomon TE, Raybould HE. Intestinal lipid inhibits emptying via CCK and a vagal capsaicin sensitive afferent pathway in rats. American Journal of Physiology 1994; 67(4tp1): G625 - 629.
- 18. Jung RT, Shetty PS. Barrant M. Callingham BA. James WPT. Role of catecholamines in hypotensive response to dieting. British Medical Journal 1979; 1:12-13.
- 19. Willet WC. Diet and Health: What should we eat? Science 1994; 264 (5158): 532 537.
- 20. Wattenburg LW. Inhibition of Neoplasia by minor dietary constituents. Cancer Research Supply 1983; 43: 2448s 2453s.
- 21. Kohlmeier L, Simonsen N, Mottus K. dietary modifiers of Carcinogens. Environmental Health Perspective 1995; 103(suppl 8): 177 184.
- 22. Stitch HF, Rosin MJ. (1984). Nutritional and Toxicological aspects of food Safety. New York. Plenum Press. pp. 1 24.
- 23. Akerele O. Nature's medicinal bounty: don't throw it away. World health Forum 1993; 14(4): 390 395.
- 24. Bhupat R. Use of Indian ethnic remedies (letter). British Medical Journal 1993; 306 (6883): 1003 1004.
- 25. WHO. Antimicrobial Resistance. Facts Sheet 2002. Retrieved 23rd September, 2007 from
- http://www.who.int/mediacentre/factsheets/fs194/en/print.ht ml
- 26. McMurry LM, Oethinger M, Levy SB. Over expression of mar A, soxS or acrAb produces resistance to triclosan in Escherichia Coli. FEMS Microbial Letter 1998; 166: 305-309.
- 27. Suller MT, Russel AD. Triclosan and antibiotic resistance in staphylococcus aureus. Journal Antimicrobial Chemotherapy 2000; 46:11-18.
- 28. Hoang TT, Scheeizer HP. Characterization of pseudomonas aeruginosa enoyl-acyl carrier protein reductase (Fabi): a target for the antimicrobial triclosian and its role in homoserine lactone synthesis, Journal of Bacteriology 1999; 181:5489-5497.
- 29. Cohen ML. Epidemiology of drug resistance: Implications for a post-antimicrobial era. Science 1992; 257:1050-1082.
- 30. Zaika LL, Kissinger JC, Wesserma AE. Inhibitory of lactic acid bacteria by herbs. Journal of Food Science 1983; 48: 1455-1459.
- 31. Shelef LA. Antimicrobial effects of spices. Journal of Food Safety 1983; 6: 29-44.
- 32. Zaika LL. Spices and herbs: Their antimicrobial acticity and its determination Journal of Food Safety 1988; 9: 97-118.
- 33. Adegoke GO, Skura BJ. Nutritional profile and antimicrobial spectrum of the spices Aframimim danielli K. shum, Plant Food for Human Nutrition 1994; 45: 175-182.
- 34. Ohlhawere MC. Antimicrobial activities of indigenous spices and condiments Kontangora Journal of Science 1996; 1 (1): 105 115.
- 35. Alonge DO, Hikko AA. Traditional methods of meat preservation in Nigeria. West African Farm Food Processing 1981; 19-21.
- 36. Okonkwo TM. "About Suya and Yaji". Journal of Food and Agriculture 1987. 1(1) 51.

- 37. Nwaopara AO, Anyanwu LC, Oyinbo CA, Anaikot IC. The Histological Changes In Pancreas Of Wister Rats Fed With Diets Containing Yaji (Local meat Sauce). Journal of Experimental and Clinical Anatomy 2004; 3(2): 44 47. 38. Betumiblog. "Kuli-Kuli is calling me to come and buy". Betumi 2006. Retrieved 27th July, 2008 from www.betumi.com
- 39. Igene JO, ID Mohammed. Consumers' attitudes towards 'suya' meat product. Annals of Borno 1983; 1.
- 40. Omojola AB. Yield and organoleptic characteristics of Suya (an intermediate moisture meat) prepared from three different muscles of a matured bull African Journal of Biotechnology (2008); 7 (13): 2254-2257.
- 41. Uzeh RE, Ohenhen RE, and Adeniji OO. Bacterial Contamination of Tsire-Suya, a Nigerian Meat Product. Pakistan Journal of Nutrition 2006; 5 (5): 458-460.
- 42. Nwaopara AO, Odike MAC, Inegbenebor U, Adoye MI. The Combined effects of excessive consumption of ginger, clove, red pepper and black pepper on the histology of the liver. Pakistan Journal of Nutrition 2007a; 6(6): 524-527.
- 43. Nwaopara AO, Odike MAC, Inegbenebor U, Nwaopara SO, Ewere GI. A comparative study on the effects of excessive consumption of ginger, clove, red pepper and black pepper on the histology of the Kidney. Pakistan Journal of Nutrition 2008a; 7(2): 287 291.
- 44. Nwaopara AO, Odike MAC, Inegbenebor U, Nwaopara SO, Ehitare IE. A Comparative study on the effects of excessive consumption of ginger, clove, red pepper and black pepper on the histology of the Heart. Electronic Journal of Biomedicine 2008b; 3 (in press).
- 45. Nwaopara AO, Odike MAC, Ikhuoriah TA, Anyanwu LC. Potential Health Hazards in Yaji: The Complex Suya Meat Sauce. Medilink Journal 2007b; 8(74): 34-38.
- 46. Jan IC. Proceedings of the 104th general meeting of the American society for microbiology.reported in birds use herbs to protect their nests, BJS Science Blog 2004; Wed 05-26.
- 47. Engel C. Wild Health: How animals keep themselves well and what we can learn from them London, Weindenfeld and Nicholson, 2002.
- 48. Hutchings MR, Athanasiaou S, Kyriazakis I, Gordon IJ. "Can animals use foraging behaviour to combat parasites? Proceedings of Nutritional Society 2003; 62 (2):361, PMID 14506883.
- 49. Dobell C. Antony van leeuwenhoek and his "little animals". New York, Dover publications Inc. 1960. pp. 141-142.
- 50. Cichewicz RH, Thorpe PA. The antimicrobial properties of chili peppers (Capsicum species) and their uses in Mayan Medicine. Journal of Ethnopharmacology 1996; 52 (2): 61-70.
- 51. Polasa, K. Ginger: Its role in Xenobiotic metabolism. ICMR Bulletin 2003; 33(6):57-63.
- 52. James ME, Nannapaneni R, Johnson MG. Identification and characterization of two bacteriocin producing bacteria isolated from garlic and ginger root. Journal of Food Protection 1999; 62: 899.
- 53. Gugnani HC, Ezenwanze EC. Antibacterial activity of extracts of ginger (Zingiber officinale) and African oil bean seed (Pentaclethora macrophylla). Journal of Communicable Diseases 1985; 17: 233.
- 54. Wood R. The whole food encyclopedia, New York, NY: Prentice-Hall Press 1988. PMID: 15220.
- 55. Foster S. Ginger-Zingiber Officinale. Monograph 2000. Retrieved 6th March, 2003, from
- http://www.stevenfoster.com/education/monograph/gin ger.html

- 56. Chrubasik S, Pittler M, Rougogalis B. Zingiberis rhizome: A comprehensive review of ginger effect and efficiency profiles. Phytomedicine 2005; 2:634-701 57. Nanir SP, Kadu BB. Effect of medicinal plant extracts on some fungi. Acta Botanica Indica 1987; 15: 170. 58. Kapoor A. Antifungal activities of fresh juice and aqueous extracts of turmeric and ginger (Zingiber officinale). Journal of Phytological Research 1997; 10: 59. 59. Meena MR. Studies on antimicrobial activity of various spices and their oils. M.Sc. Thesis: Indian Agricultural Research Institute, New Delhi 1992.
- 60. Pruthi JS. Spices and condiments: Chemistry, Microbiology, Technology. New York Academic Press 1980.
- 61. Wendakdoon CN, Sakaguchi M. Combined effect of sodium chloride and clove on growth and biogenic amine formation of Enterobacter aerogenes in mackere muscle extracts. Journal of Food Protection 1993; 56, 410-413. 62. Ting WTE, Diebel KE. Sensitivity of listeria
- monocytogenes to spices at two temperatures. Journal of Food Safety 1992; 12, 129-137.
- 63. Aureli P, Costentini A, Zolea S. Antimicrobial activity of some plant essential oils against Listeria monocytogenes. Journal of Food Protection 1992; 55, 344-348.
- 64. Smith-Palmer A, Stewart J, Fyfe L. Antimicrobial properties of plant essential oils and essences against five important food-borne pathogens. Lett Appl. Microbial 1998; 28, 118-122.
- 65. Stecchini ML, Sarais L, Gravedoni P. Effect of essential oils on Aeromonas hydrophila in a culture medium and in cooked pork. Journal of Food Protection 1993; 56, 406-409. 66. Bara MTF, Vanetti MCD. Antimicrobial effect of spices on the growth of Yersinia enterocolitica. Journal Herbs Spices Medicinal Plants 1995; 3: 51-58.
- 67. Degre R, Syvester M. Effect of butylated hydroxyanisole on the cytoplasmic membrane of staphylococcus aureus wood 46. Journal of Food Protectection 1983; 46:206-209. 68. Allen SB, Baron EJ. Clostridrium. In: Balows, A. et al. (Eds), Manual of Clinical Microbiology. 5th ed. Washington, DC: America Society for Microbiology 1991; pp. 505-521.
- 69. Molina-Torres, J. et al. 1999. Antimicrobial properties of alkamides present in flavouring plants traditionally used in Mesoamerica: affinin and capsaicin. Journal of Ethnopharmacology 64(3): 241-248.
- 70. Perez C, Anesinic C. Antibacterial activity of alimentary plants against Staphylococcus aureus growth. American Journal Clinical Medicine 1994; 22:169-174.
- 71. Singh G, Marimuthu P, Murali HS Bawa AS. Antioxidative and antibacterial potential of essential oils and extracts isolated from various spice materials. Journal of Food Safety 2005; 25(2):130.
- 72. Adebesin, AA, Saromi, OT, Amusa, NA and Fagade, SO. Microbiological quality of some groundnut products hawked in Bauchi, a Nigerian City. The Journal of Food Technology in Africa 2001; 6(2): 53-55.
- 73. Bernier G, Morin, M, Marsolais G. A Generalized Inclusion Body Disease in the Budgerigar (Melopsittacus undulatus) Caused by a Papovavirus-Like Agent. Avian Diseases 1981. 25(4) 1083-1092.
- 74. Nickeleit V, Hirsch HH, Binet IF et al. Polyomavirus infection of renal allograft recipients: from latent infection to manifest disease. Journal of American Society of Nephrology 1999. 10: 1080-1089.
- 75. Nickeleit V, Hirsch, HH, Zeiler M, Gudat F, Prince, O, Thiel G, Mihatsch MJ.
- BK-virus nephropathy in renal transplants-tubular necrosis,

Antimicrobial Potentials of Yaji-Spices: The Constituents of a Complex Nigerian Suya Meat Sauce Inducing Histological Investigations

- MHC-class II expression and rejection in a puzzling game. Nephrol Dial Transplant 2000; 15: 324-332.
- 76. Drachenberg CB, Beskow CO, Cangro CB et al. Human polyoma virus in renal allograft biopsies: morphological findings and correlation with urine cytology. Human Pathology 1999; 30: 970-977.
- 77. Grieve MA. A Modern Herbal: Dover publications, New York 1971.
- 78. Saber MS. Antimicrobial substances in certain members of Solanaceae IV. Detection of active principles in pepper plant Zentralbl Bakteriol Parasitenkd Infektionskr Hyg 1976; 131(2): 110.
- 79. Kurokawa M. Hozuni T, Basnet P, Nakano M, Namba T,
- Kawana T, Shiraki T. Purification and Characterization of eugenine as anti herpes virus compound from japonicum and syzygium aromaticum. Journal of Pharmacology and Exp Therap 1998. 284 (2): 728 735.
- 80. Amaechi BT, Higham SM, Edgar WM. Techniques for the production of dental eroded lesions in vitro. J Oral Rehabil 1999 Feb;26(2):97-102. PMID:12580
- 81. Ghelardini C, Galeotti N, Di Cesare, ML. Local anaesthetic activity of bata-caryophyllene. Farmaco 2001; 56 (5-7): 387-9, PMID: 12570.
- 82. Kalemba D, Kunicka A. Antibacterial and antifungal properties of essential oils. Curr Med Chem 2003; 10 (10): 813-29.

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