

Hematological and biochemical studies on filariasis of dogs

M Hashem, A Badawy

Citation

M Hashem, A Badawy. *Hematological and biochemical studies on filariasis of dogs*. The Internet Journal of Veterinary Medicine. 2007 Volume 4 Number 2.

Abstract

The present work was done on eighty two stray dogs to be investigated for filariasis via hematological and serum chemistry profiles of naturally infested dogs. Out of the examined dogs, 14 (17.1%) were infested with *Dipetalonema reconditum*, 12 (85.7%) of them were males and 2 dogs (14.3%) were females. Microfilariae appeared as a snake like with a rapidly, forward movement across the microscopic field in wet smear while in Giemsa stained smears showed a coiled or twisted appearance. Hematological studies revealed hemolytic anemia associated with low erythrocyte counts, hemoglobin concentration and hematocrit value. A marked increase in erythrocyte sedimentation rate, reticulocyte, thrombocyte, total and differential leucocytic counts were encountered, in comparison with the control group.

Biochemical analysis of sera from infested dogs showed a significant changes in the determined parameters used for evaluation of liver and kidney functions.

It could be concluded that infestation of dogs with filariasis induced a hemolytic anemia, with disturbance in the liver and kidney functions.

INTRODUCTION

Filariasis is one of the most important parasitic diseases caused by the filaroid nematodes with a world wide distribution and affects both man, animals and birds. In Egypt, while many previous studies on herbivorous animal filariasis were conducted both in Sharkia and other provinces throughout the country (^{1,2,3,4,5,6}) few studies dealt with filariasis of dogs (^{6,7,8,9}) and no records of filariasis in this animal species in Sharkia province were documented. On the other hand, local studies on hemoparasites in dogs with particular relation to hematological and biochemical dimensions are limited (¹⁰).

From this point of view and since filariasis of dogs (dirofilariasis) represent a public health hazards to man (^{11,12}), this study was conducted to investigate the dogs for filariasis and to study the blood cellular and biochemical changes in naturally filariasis infested dogs.

MATERIALS AND METHODS

DOGS AND BLOOD SAMPLES

Blood samples were collected from 82 middle aged stray dogs and examined directly for filariasis. For haematological and biochemical analysis, blood samples were collected from ten microfilaraemic dogs proved free from internal and

external parasites through naked eye, blood and faecal examinations. As a control group, five dogs of a comparable age were treated with Praziquantel (5 mg/kg body weight, orally) and Ivermectin (1 ml/50 kg body weight, subcutaneously) and proved to be free from internal and external parasites through repeatedly naked eye, faecal and blood examinations over a period of three months post treatment were used. Blood samples for haematological and biochemical analysis were divided into two portions as following: The 1st portion (5ml) put in clean dry test tubes containing anticoagulants as sodium citrate 3.8% for determination of erythrocyte sedimentation rate, dipotassium salt of EDTA for studies of erythrogram and leucogram, and ammonium oxalate 1% for platelet counts. The 2nd portion (6ml) put in plain centrifuge tubes, left undisturbed for clotting of the blood and the clear straw-coloured serum was carefully separated after centrifugation at 3000 r.p.m. for 15 minutes and kept in the deep freezing at -200C until subsequent biochemical analysis.

PARASITOLOGICAL STUDIES

Wet smears, modified Knott technique (¹³) as well as Giemsa stained blood films were used to investigate dogs for microfilariae. The microfilariae were measured using a calibrated eye micrometer and photographed using Leitz

microscope (Germany) and Canon digital photo camera (Japan). To study the microfilarial periodicity, blood samples were collected every three hours from three microfilaraemic dogs and used to investigate the day periodicity of microfilariae using the technique of Ezzat and Tadros (14). In brief, 0.5 ml of freshly collected blood was added to 1.5 ml of 2% glacial acetic acid in distilled water tinged with gentian violet. After thorough mixing, the tubes were left for 5 min. then the number of microfilariae was counted in 0.1 ml of the mixture and multiplied by 40 to give the number of microfilariae in one ml blood.

HEMATOLOGICAL ANALYSIS

The hematological parameters included erythrocyte sedimentation rate (ESR), red blood cell count (RBCs), hemoglobin concentration (Hb), packed cell volume (PCV), reticulocyte count (using Brilliant cresyl blue stained film), platelet count as well as total and differential leucocytic counts were performed using standard techniques as described by Feldman et al. (15). The blood indices included mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH) and mean corpuscular hemoglobin concentration (MCHC) were calculated.

BIOCHEMICAL ANALYSIS

Serum samples were colorimetrically analyzed for the activities of alanine aminotransferase (ALT), aspartate aminotransferase (AST), bilirubin (total, direct & indirect), glucose, total proteins, albumin, globulins (calculated as the difference between total proteins and albumin) as a biochemical indicators for liver function. Serum levels of urea nitrogen, creatinine, inorganic phosphorus, calcium, sodium, potassium and magnesium were used for evaluation of kidney function. All the biochemical analyses were measured using the determination methods according to manufacturer instructions (kits from Bio-merieux, France).

STATISTICAL ANALYSIS

The obtained data in this study were computed and statistically analyzed using student's "t" test according to Tamhane and Dunlop (16).

RESULTS

PREVALENCE OF FILARIASIS OF DOGS

Out of 82 examined dogs, 14 dogs (17.1%) were proved to be infested with *Dipetalonema reconditum* according to the microfilarial identification. Out of 14 infested dogs, 12 dogs (85.7%) were males and 2 dogs (14.3%) were females.

MORPHOLOGY OF THE MICROFILARIA

In wet blood smears, the microfilariae appeared as a snake like with a rapidly, forward movement across the microscopic field. Stained microfilariae appeared coiled or twisted to various degrees (plate 1, A). The microfilarial length varied from about 250 – 260 µm (aver. 255± 2.4 µm), while the diameter varied from about 3.5 – 4.5 µm (aver. 4± 0.24 µm). The anterior end of the microfilariae devoid from nuclei to a distance about 7 – 8 µm (aver. 7 ± 0.45 µm), (plate 1, B). The nerve ring and excretory pore located at about 28 – 32 µm (aver. 30± 0.68 µm) and 40 – 44 µm (aver. 42± 0.84 µm) from the anterior end, respectively. The anal pore located at about 60 – 70 µm (aver. 65±0.98 µm) from the tail end which showed mostly a hooked appearance (plate 1, C).

Figure 1

Plate 1: The microfilaria of , Giemsa stained. (A): The whole microfilaria (Bar = 30 µm), (B): The anterior end of the microfilaria with no nuclei (Bar = 7 µm), (C): The posterior end of the microfilaria showing a characteristic hooked tail (Bar = 10 µm).

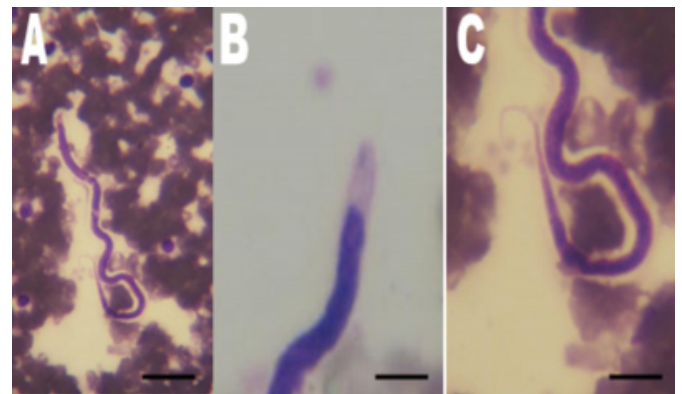
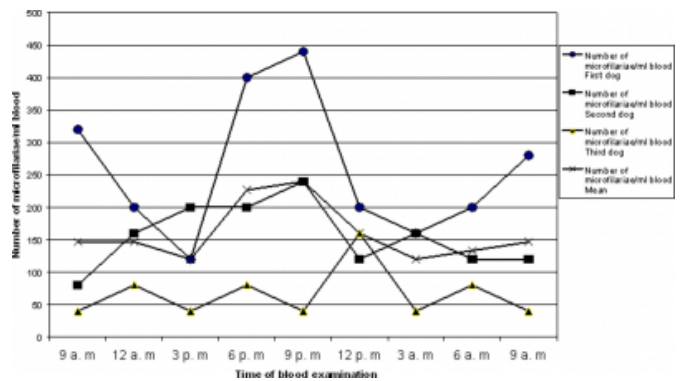


Figure 2

Figure 1: , a summer day microfilarial periodicity.



MICROFILARIAL PERIODICITY

As shown in Fig. 1, the number of microfilariae increased

significantly in the peripheral blood toward the evening (nocturnal periodicity) and peaked between 6 – 9 p. m.

BLOOD CELLULAR FINDINGS

Blood cellular analysis of *Dipetalonema reconditum* infested dogs revealed a significant reduction in RBCs counts, Hb content, PCV value and increase in reticulocyte count, MCV, MCH with a decrease in MCHC, indicating the presence of regenerative anemia of macrocytic hypochromic type. The values of ESR, reticulocyte, thrombocyte, total and differential leucocytic counts were significantly increased, in comparison with the control group (table 1).

BIOCHEMICAL FINDINGS

Liver function tests of sera from infested dogs showed a significant ($P \leq 0.01$) increase in the serum activities of alanine aminotransferase (ALT) and aspartate aminotransferase (AST), serum bilirubin (total & indirect), total proteins, globulins and a decrease ($P \leq 0.05$) in the serum values of glucose and albumin, with insignificant change in the serum direct bilirubin, when compared with control (table 2).

Kidney function tests in infested dogs revealed a significant increase in serum urea nitrogen ($P \leq 0.01$), creatinine, inorganic phosphorus, potassium and a decrease in the serum calcium and sodium levels ($P \leq 0.05$) while, the serum magnesium level showed insignificant change, comparatively with control (table 3).

Figure 3

Table 1: Blood cellular parameters in the control and infested dogs with (Mean values \pm SD).

Parameters	Normal dogs	Infested dogs
Erythrocyte sedimentation rate (mm/h)	2.45 \pm 0.12	3.73 \pm 0.16*
Red blood cell count ($\times 10^6/\mu\text{l}$)	6.5 \pm 0.45	3.4 \pm 1.12**
Hemoglobin concentration (g/dl)	16.0 \pm 1.12	9.5 \pm 1.12**
Packed cell volume (%)	40.0 \pm 2.42	28.0 \pm 1.12**
Mean corpuscular volume (fl)	61.53 \pm 5.6	82.35 \pm 1.12*
Mean corpuscular hemoglobin (Pg)	24.61 \pm 3.15	27.94 \pm 1.12*
Mean corpuscular hemoglobin concentration (%)	40.0 \pm 4.37	33.92 \pm 1.12*
Reticulocytes (%)	1.2 \pm 0.02	2.5 \pm 1.12**
Platelets ($\times 10^3/\mu\text{l}$)	450 \pm 17.61	760 \pm 17.61**
Total leukocytic count ($\times 10^3/\mu\text{l}$)	8.5 \pm 1.12	16.6 \pm 1.12**
Segmented Neutrophils ($\times 10^3/\mu\text{l}$)	4.0 \pm 0.2	6.5 \pm 0.2*
Band neutrophils ($\times 10^3/\mu\text{l}$)	0.1 \pm 0.001	0.4 \pm 0.01**
Lymphocytes ($\times 10^3/\mu\text{l}$)	3.4 \pm 0.15	5.4 \pm 0.15*
Monocytes ($\times 10^3/\mu\text{l}$)	0.7 \pm 0.05	2.5 \pm 1.35**
Eosinophils ($\times 10^3/\mu\text{l}$)	0.3 \pm 0.01	1.8 \pm 1.25**

*Significant at probability ≤ 0.05 **Significant at probability ≤ 0.01

Figure 4

Table 2: Liver function tests in the control and infested dogs with (Mean values \pm SD).

Parameters	Normal dog	Infested dog
Alanine aminotransferase (U/l)	55.24 \pm 2.05	120.66 \pm 4.30**
Aspartate aminotransferase (U/l)	80.0 \pm 1.4	150.0 \pm 5.33**
Total bilirubin (mg/dl)	0.4 \pm 0.06	1.05 \pm 0.2**
Direct bilirubin (mg/dl)	0.15 \pm 0.01	0.15 \pm 1.12
Indirect bilirubin (mg/dl)	0.25 \pm 0.03	0.90 \pm 0.02**
Glucose (mg/dl)	86.0 \pm 1.76	40.8 \pm 1.12*
Total protein (g/dl)	7.00 \pm 0.75	8.95 \pm 0.16*
Albumin (g/dl)	3.70 \pm 0.55	3.0 \pm 0.04*
Globulins (g/dl)	3.30 \pm 0.61	5.95 \pm 0.32*

*Significant at probability ≤ 0.05 **Significant at probability ≤ 0.01

Figure 5

Table 3: Kidney function tests in the control and infested dogs with (Mean values \pm SD).

Parameters	Normal dog	Infested dog
Urea nitrogen (mg/dl)	25.0 \pm 1.77	73.92 \pm 2.04**
Creatinine (mg/dl)	1.2 \pm 0.02	1.5 \pm 0.02*
Inorganic phosphorus (mg/dl)	2.8 \pm 0.02	3.5 \pm 1.12*
Calcium (mg/dl)	10.5 \pm 1.1	8.4 \pm 1.12*
Sodium (mEq/l)	141.4 \pm 12.5	125.8 \pm 17.61*
Potassium (mEq/l)	4.5 \pm 1.12	6.6 \pm 0.72*
Magnesium (mg/dl)	1.9 \pm 0.2	2.0 \pm 0.2

*Significant at probability ≤ 0.05 **Significant at probability ≤ 0.01

DISCUSSION

In the present study, a survey was conducted to investigate the dogs for filariasis, as well as the blood cellular and biochemical changes in naturally infested dogs. Out of 82 examined dogs, 14 (17.1%) were infested with *Dipetalonema reconditum*. A nearly similar infestation rates of dogs with *Dipetalonema reconditum* were also reported, in which 22.6% of dogs were infested in Brazil₍₁₇₎ and 15.9% infestation rate was reported in dogs from South Italy₍₁₈₎. While, lower infestation rates with *Dipetalonema reconditum* were also recorded in dogs in Egypt and other countries, where 0.063% of dogs from Abu Rawach, Giza, Egypt proved to be infested (9), 1.0% of dogs in Spain were infested₍₁₉₎, less than 0.5% infestation rate in the State of Washington₍₂₀₎ and 6% in Western Sicily, Italy₍₂₁₎. Reasons for these differences in infestation rates in these studies may be attributed to the locality, distribution and prevalence of the arthropod vectors of this parasite such as fleas, lice and ticks, which in great part affected by the different climatic conditions in these regions as well as the methods of examination of dogs for filariasis. High infestation rate was recorded in male dogs (85.7%) than in females (14.3%). Similar results were stated by Falls and Platt₍₂₂₎ and Amer₍₆₎. This is might be returned to hormonal effect on

susceptibility of dogs to infestation.

Regarding the observed characteristic morphological features of *Dipetalonema reconditum* microfilariae in this study, there was no contradiction with the previous descriptions (23,24,25,26). Concerning with the microfilarial periodicity of *Dipetalonema reconditum*, this study showed a nocturnal periodicity of the microfilariae and peaked between 6 – 9 p. m. These results were to some extent in agreement with the results of Newton and Wright (13) who reported a nocturnal periodicity of microfilariae of *Dipetalonema reconditum* with two peaks at 6.0 p. m and 12 p. m. and Korkejian and Edeson (27) who noticed nocturnal periodicity of microfilariae of *Dipetalonema reconditum*. Also, Amer (9) observed increase the number microfilariae of this parasite in peripheral blood of infested dogs between 6.30 p. m – 10.30 p. m during the different seasons of the year.

Concerning the hematological results in the present work, a regenerative anemia of macrocytic hypochromic type associated with a reduction in the RBCs count, Hb concentration and PCV value were recorded, with an increase of ESR, reticulocyte, thrombocyte, total and differential leucocytic counts. The macrocytosis and hypochromasia were due to reticulocytosis that seen in the infested dogs with microfilariae. The present anemia may be attributed to the hemolysis of RBCs as a result of destructive motility of microfilaria as reported by Ishihara et al. (28) and (29) and Kitagawa et al. (30) who showed a severe intravascular hemolysis with a significant reduction of RBCs count and Hb concentration in dogs with dirofilariasis. Ziegler et al. (31) found intravascular hemolytic anemia (macrocytic up to 80 days after infection, subsequently normocytic and hypochromic), accompanied by reticulocytosis in the rodent, *Mastomys natalensis*, infested with *Litomosoides carinii*. Similar findings were obtained by previous authors (10,32,33,34). Moreover, Sharma and Joshi (35) showed a decrease in the erythrogram of microfilariae infested cattle. Reifur et al. (17) reported a significant macrocytic anemia in dogs infested with three different microfilariae: *Dirofilaria immitis*, *D. reconditum*, and the third (mf3) were not identified. The latter authors mentioned that *D. reconditum* was the species with the highest prevalence (22.6%), while *Dirofilaria immitis* was 5.47%. Our results disagree with Anuchai et al. (36) who found moderate microcytic anemia and severe thrombocytopenia in 7 dogs infested with dirofilariasis, ehrlichiosis, and babesiosis. The difference may be

attributed to the complicated infestations in these dogs, while in our study; we found only *D. reconditum* microfilariae.

The higher ESR value in infested animals may be due to the anemia. It may also be due to auto-agglutination that is observed in this disease during infection. The increase in ESR has been observed in many other diseases where autoagglutination of red blood cells takes place as in malaria and tuberculosis (37). Similar results were obtained in canine with dirofilariasis (10), in microfilariae-infested cattle (35), in haemoparasitized camels with *Trypanosoma evansi* and *Dipetalonema evansi*, (38) and in an owl with microfilaraemia (39).

Thrombocytosis observed in hemoparasitised dogs could be related to the hemolytic anemia (10, 40). On contrary, thrombocytopenia was obtained by Rawlings (41) and Anuchai et al. (34) in dogs infested with *Dirofilaria immitis*.

The leukogram revealed a marked leucocytosis with neutrophilia, eosinophilia, lymphocytosis and monocytosis. The higher blood neutrophil and monocyte counts were for the phagocytic removal of tissue breakdown products or microfilariae. Similarly, Paltrinieri et al. (42) showed neutrophilic leucocytosis in dogs with dirofilariasis. The observed eosinophilia was due to sensitivity to the foreign protein of a parasite which may be a part of an immune phenomenon (15). The lymphocytosis which develops in dogs infested with blood parasite is presumably due to intense antigenic stimulations which increase the demands for lymphocytes to be transformed into plasma cells for antibodies production. Yamagata et al. (43) found lymphocytosis with increases in IgE values in dogs experimentally co-infested with *Dirofilaria immitis* and *Ancylostoma caninum*. The authors mentioned that parasitic nematodes that undergo blood and tissue migrations increased IgE and IgG values. The results of the leukogram were in agreement with the findings of others (10, 34, 35, 41, 44).

Concerning the biochemical results, an increase in the serum enzyme activities (ALT & AST), serum bilirubin (total & indirect) and a decrease in the serum glucose level were observed in the dogs infested with *D. reconditum*, when compared with the non infested one. The increased serum enzymes and hypoglycemia demonstrated in microfilariaemic dogs suggested liver dysfunction secondary to circulatory disturbance. In addition, the hypoglycemia was attributed to glucose consumption by the *Dipetalonema viteae* and *B. pahangi* parasites (45). The hyperbilirubinemia

(total & indirect) may be attributed to hemolytic anemia with resultant hemolytic jaundice. The obtained results were in harmony with earlier findings (10, 31, 34, 36, 38, 46).

Protein profile of serum samples showed an increase in the total protein and globulins concentration with a decrease in the albumin values in the infested dogs with microfilariae comparatively with non-infested one. The observed hyperproteinemia can be attributed on the one hand to an increase in the γ -globulin concentration in response to the parasitic antigens and on the other had to a release of hemoglobin from destructed erythrocytes (47). The obtained hypoalbuminemia probably corresponds to the degenerative changes in the haemoparasitized organs (mainly liver). Similar results have been reported previously (32, 35, 38, 48).

The significantly higher serum urea nitrogen, creatinine, inorganic phosphorus, potassium and lower serum calcium and sodium levels in infested dogs than in non infested one might result from more severe kidney dysfunction, metabolic acidosis, as well as intravascular hemolysis (30, 32, 36).

In conclusion, a hemolytic anemia with disturbance in the liver and kidney functions were the main results in canine filariasis, caused by *Dipetalonema reconditum*.

ACKNOWLEDGMENTS

We thank all the staff members of Clinical Pathology and parasitology Departments, Faculty of Veterinary Medicine, Zagazig University, for their valuable help and continuous facilities which made the work calmly achieved.

References

1. El Seify MA, El Askalany MA, Rashad SM. Biochemical changes in serum of buffaloes naturally infected with microfilariae in Beni-seuf Governorate. Egypt J Comp Pathol Clin Pathol 1990; 3 (1): 27 - 36.
2. Sakla AA. Parasitological studies on filariasis in Assiut Governorate. Assiut Med J 2000; 24 (1): 37 - 46.
3. El-Massry AA, Derbala AA. Evidence of *Onchocerca fasciata* (Filaroidea: Onchocercidae) in camels (*Camelus dromedaries*): Prevalence, nodular lesions appearance and parasite morphology. Vet Parasitol 2000; 88 (3-4): 305 - 312.
4. Arafa MI. Studies on ecto and endoparasites of equines in Assiut Governorate. PhD Thesis, Fac Vet Med Assiut University, 2002.
5. Mahran ON. Some studies on blood parasites in camels (*Camelus dromedaries*) at Shalatin city, Red Sea Governorate. Assiut Vet Med J 2004; 50(102):172- 184.
6. Bahnass MM. Studies on filariasis in some farm animals. MV Sc Thesis, Fac Vet Med, Zagazig University, 2005.
7. Fahmy LS. Some surgical problems caused by filariae in domesticated animals. MVSc Thesis, Fac Vet Med, Cairo University, 1972.
8. Ahmed BA, Amer OH, Fayek SA. New record for *Dirofilaria repens* (Raillet and Henry, 1911). Adult worm and its pathogenesis in naturally infested dogs in Egypt. Zagazig Vet Med J 1986; 16 (1): 89 - 98.
9. Amer OH. Some studies on filaria in dogs. Ph. D. Thesis, Fac. Vet. Med., Zagazig University, 1986.
10. Sharma MC, Pachauri SP. Blood cellular and biochemical studies in canine dirofilariasis. Vet Res Commun 1982; 5 (3): 295 - 300.
11. Gorezis S, Psilla M, Asproudis I, et al. Intravitreal dirofilariasis: a rare ocular infection. Orbit 2006; 25 (1): 57 - 59.
12. Sathyan P, Manikandan P, Bhaskar M, et al. Subtenons infection by *Dirofilaria repens*. Indian J Med Microbiol 2006; 24(1): 61-62.
13. Newton WI, Wright WH. The occurrence of dog filariid other than *Dirofilaria immitis* in the United States. J Parasitol 1956; 42: 246 - 248.
14. Ezzat MAE, Tadros G. A simple method for counting microfilariae in blood of animals. Agr Res Rev 1958; 36 (4): 38 - 44.
15. Feldman BF, Zinkl JG, Jain NC. Schalm's Veterinary Hematology. 5th Ed., Lippincott Williams and Wilkins. A Walters Company. Philadelphia, Baltimore, New York, London, Buenos Aires, Hong Kong, Sydney, Tokyo, 2000.
16. Tamhane AC, Dunlop DD. Statistical and data analysis from elementary to intermediate. Upper Saddle River, USA, 2000.
17. Reifur L, Thomaz-Soccol V, Montiani F. Epidemiological aspects of filariasis in dogs on the coast of Parana state, Brazil: with emphasis on *Dirofilaria immitis*. Vet Parasitol 2004; 122 (4): 273 - 286.
18. Cringoli G, Rinaldi L, Capelli G. A prevalence survey and risk analysis of filariasis in dogs from the Mt. Vesuvius area of southern Italy. Vet Parasitol 2001; 102 (3): 243 - 252.
19. Ortega-Mora LM, Gomez-Bautista M, Rojo-Vazquez F, Rodenas A, Guerrero JA. Survey of the prevalence of canine filariasis in Spain. Prev Vet Med 1991; 11: 63 - 68.
20. Theis J, Stevens F, Law M. Distribution, prevalence, and relative risk of filariasis in dogs from the State of Washington (1997-1999). J Am Anim Hosp Assoc 2001; 37 (4): 339 - 347.
21. Giannetto S, Pampiglione S, Santoro V, Virga A. Research of canine filariasis in Trapani province (western Sicily). Morphology on SEM of male *Dirofilaria repens*. Parasitologia 1997; 39 (4): 403 - 405.
22. Falls RK, Platt TR. Survey of heart worm, *Dirofilaria immitis*, and *Dipetalonema reconditum* (Nematoda, Filaroidea) in dogs from Virginia and North Carolina. Am. J Vet Res 1982; 43 (4): 738 - 739.
23. Nelson GS. *Dipetalonema reconditum* (Grassi, 1889) from the dog with a note on its development in the flea, *Ctenocephalides felis* and the louse, *Heterodoxus spiniger*. J Helminthol 1962; 3: 297 - 308.
24. Kelly JD. Detection and differentiation of microfilariae in canine blood. Aus Vet J 1973; 49 (1): 23 -27.
25. Watson AD, Testoni FJ, Porges WL. A comparison of microfilariae isolated from canine blood by the modified Knott test and a filter method. Aus Vet J 1973; 49 (1):28 - 30.
26. Soulsby E.J.L. Helminthes, Arthropodes and Protozoa of Domesticated animals. 7th ed., Bailliere Tindall, London, 1982.
27. Korkejian A, Edeson JF. Studies on naturally occurring filarial infections in dogs in Lebanon. I. *Dipetalonema reconditum*. Ann Trop Med Parasitol 1978;72 (1): 65 - 78.
28. Ishihara K, Kitagawa H, Ojima M, Suganuma Y. Clinicopathological studies on canine dirofilarial hemoglobinuria. Nippon Juigaku Zasshi 1978;40 (5):

525-537.

29. Ishihara K, Kitagawa H, Yokoyama S, Ohashi H. Studies on hemolysis in canine dirofilarial hemoglobinuria. Lipid alterations in blood serum and red cell membrane. *Nippon Juigaku Zasshi* 1981; 43 (1): 1 - 11.
30. Kitagawa H, Sasaki Y, Ishihara K. Clinical studies on canine dirofilarial hemoglobinuria: measured and calculated serum osmolalities and osmolar gap. *Nippon Juigaku Zasshi* 1989; 51 (4): 703 - 710.
31. Ziegler C, Käufer-Weiss I, Zahner H. On the pathogenesis of anaemia and leukopenia in filarial (*Litomosoides carinii*) infection of *Mastomys natalensis*. *Zentralbl Veterinarmed B* 1991;38 (2):123 - 134.
32. Kitagawa H, Kitoh K, Ohba Y, et al. Comparison of laboratory test results before and after surgical removal of heartworms in dogs with vena caval syndrome. *J Am Vet Med Assoc* 1998; 213 (8): 1134-1136.
33. Nielsen NO, Simonsen PE, Magnussen P, Magesa S, Friis H. Cross-sectional relationship between HIV, lymphatic filariasis and other parasitic infections in adults in coastal northeastern Tanzania. *Trans Royal Soc Trop Med Hyg* 2006; 100 (6): 543 - 550.
34. Anuchai N, Morakot K, Sonthaya T, Somporn T, Siram SA. Retrospective study of the clinical hematology and the serum biochemistry tests made on canine dirofilariasis cases in an animal hospital population in Bangkok, Thailand. *Research in Veterinary Science* 2007; 82 (3): 364 - 369.
35. Sharma M, Joshi C. Serum mineral and haemato-biochemical profile of microfilariae infested cattle in India: Its effects on production and therapy. *Asian-Australasian journal of animal* 2002; 15 (3): 357 - 365.
36. Anuchai N, Sukullaya A, Somporn T, Siram S, Morakot K. Canine dirofilariasis and concurrent tick-borne transmitted diseases in Bangkok, Thailand. *J Comp Clin Pathol* 2006; 15 (4): 249 - 253.
37. Hagan WA, Bruner DW. *The Infectious Diseases of Domestic Animals*. Comstock Pub. Assoc., A divisional Cornell Univ. Press, Ithaca, New York, USA, 1991.
38. Shafqaat A, Butt AA, Muhammad G, Athar M, Khan MZ. Haemato-biochemical studies on the haemoparasitized camels. *Int J Agriculture & Biol* 2004; 6 (2): 331-334.
39. Bedin M, Petterino C, Gallo E, Selleri P, Morgante M. Clinical Pathological Findings in an Owl (*Athene noctua*) with Microfilaraemia in Italy. *J Vet Med A Physiol Pathol Clin Med* 2007; 54 (3):128 - 130.
40. Makiya K. Recent increase of human infections with dog heartworm *Dirofilaria immitis* in Japan. *Parassitologia* 1997; 39 (4): 387 - 388.
41. Rawlings CA. Clinical laboratory evaluations of seven heartworm infested beagles: during disease development and following treatment. *Cornell Vet* 1982; 72 (1): 49 - 56.
42. Paltrinieri S, Sartorelli P, De Vecchi B, Agnes F. Metabolic findings in the erythrocytes of cardiopathic and anaemic dogs. *J Comp Pathol* 1998; 118 (2): 123 - 133.
43. Yamagata GR, Gershwin LJ, Wong MM. Diethylcarbamazine-induced *Dirofilaria immitis* larval death, as indicated by immunoglobulin E concentration, in dogs with concurrent *Ancylostoma caninum* infection. *Am J Vet Res* 1995; 56 (2): 174 - 178.
44. Gossett KA, Root CR, Cleghorn B, Church GE, Turk MA. Effects of heartworm and intestinal parasitic infections on hematology and peripheral lymph node cytology in Louisiana dogs. *Vet Clin Pathol* 1987; 16 (4): 97 - 101.
45. Court JP, Martin-Short M, Lees GM. A comparison of the response of *Dipetalonema viteae* and *Brugia pahangi* adult worms to antifilarial agents. *Trop Med Parasitol* 1986; 37 (4): 375 - 380.
46. Kitagawa H, Kitoh K, Iwasaki T, Sasaki Y. Comparison of laboratory data in dogs with heartworm caval syndrome surviving and non surviving after surgical treatment. *J Vet Med Sci* 1997; 59 (7): 609 - 611.
47. Moustafa AM, Agag B, Esmat M, Selim AM. Studies on filariasis in Egyptian buffaloes. III. Clinical observations and electrophoretic patterns in sera of naturally infested buffaloes with microfilaria before and after treatment with stipophon. *Zagazig Vet J* 1991; 19: 583 - 595.
48. Safwat MS, El-Abdin Y Z. Some biochemical studies on the serum of infested and non-infested camels with *Dipetalonema evansi*. *Egyptian J Vet Sci* 1982; 19: 141 - 145.

Author Information

Mohamed Hashem

Dept. of Clinical Pathology, Faculty of Vet. Med., Zagazig University

Ahmed Badawy

Dept. of Parasitology, Faculty of Vet. Med., Zagazig University